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## Lipids as Tumoricidal Components of Human Alpha-lactalbumin Made Lethal to Tumor Cells (HAMLET); Unique and Shared Effects on Signaling and Death.

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*Published in:*  
Journal of Biological Chemistry

*DOI:*  
[10.1074/jbc.M113.468405](https://doi.org/10.1074/jbc.M113.468405)

2013

[Link to publication](#)

### *Citation for published version (APA):*

Ho Cs, J., Storm, P., Rydström, A., Bowen, B., Alsin, F., Sullivan, L., Ambite, I., Mok, K. H., Northen, T., & Svanborg, C. (2013). Lipids as Tumoricidal Components of Human Alpha-lactalbumin Made Lethal to Tumor Cells (HAMLET); Unique and Shared Effects on Signaling and Death. *Journal of Biological Chemistry*, 288(24), 17460-17471. <https://doi.org/10.1074/jbc.M113.468405>

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10

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Lipids as Tumoricidal Components of Human Alpha-lactalbumin Made Lethal to Tumor Cells (HAMLET); Unique and Shared Effects on Signaling and Death

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\*Running title: *Lipids, HAMLET and cell death*

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**Keywords:** HAMLET; oleate; oleic acid; tumor cell death; gene expression; lipid metabolism

**Background:** HAMLET is a broadly tumoricidal complex of partially unfolded  $\alpha$ -lactalbumin and oleic acid, whose structural and functional contributions to HAMLET remain largely undefined.

**Results:** The protonation state and function of the lipid are elucidated.

**Conclusion:** HAMLET and oleate possess unique and overlapping effects on tumor cells.

**Significance:** Unfolded proteins, like  $\alpha$ -lactalbumin form novel functional entities with deprotonated fatty acids.

**showed weak effects on ion fluxes and gene expression. Unlike HAMLET, which causes metabolic paralysis, fatty acid metabolites were less strongly altered. The functional overlap increased with higher oleate concentrations (500uM). Cellular responses to OA were weak or absent, suggesting that deprotonation favors cellular interactions of fatty acids. Fatty acids may thus exert some of their essential effects on host cells when in the deprotonated state and when presented in the context of a partially unfolded protein.**

## SUMMARY

Long-chain fatty acids (LCFAs) are internalized by receptor-mediated mechanisms or receptor-independent diffusion across cytoplasmic membranes and are utilized as nutrients, building blocks and signaling intermediates. Here we describe how the association of LCFAs to a partially unfolded, extracellular protein can alter the presentation to target cells and cellular effects. HAMLET (Human  $\alpha$ -lactalbumin made lethal to tumor cells) is a tumoricidal complex of partially unfolded  $\alpha$ -lactalbumin and oleic acid (OA). As OA lacks independent tumoricidal activity at concentrations equimolar to HAMLET, the contribution of the lipid has been debated. We show by natural abundance <sup>13</sup>C NMR that the lipid in HAMLET is deprotonated and by chromatography that oleate rather than oleic acid is the relevant HAMLET constituent. Compared to HAMLET, oleate (175  $\mu$ M)

Long-chain fatty acids (LCFAs) are essential cellular components, serving as nutrients, membrane constituents, signaling molecules and precursors for prostaglandins and other crucial bioactive substances (1). Examples of their effects include modifications of enzymatic function, gene expression, synaptic transmission and metabolism (2-5). Dysregulated LCFA function is associated with numerous medical disorders, including infection, inflammation, atherosclerosis and cancer (6-9). To fulfill these diverse functions, fatty acids engage with cell membranes and specific fatty acids are taken up from the circulation. A variety of membrane interaction mechanisms have been characterized, but many aspects remain unclear. Fatty acids cross lipid bilayers via flip-flop mechanisms or specific protein-receptor interactions. Receptors identified in caveolae/lipid rafts (10) include FAT (CD36), caveolin-1 and interacting cytosolic fatty acid binding proteins (FABs), which bind anionic phospholipids as well as lipids modified by peroxidation (11,12).

HAMLET (human  $\alpha$ -lactalbumin made lethal to tumor cells) is a complex of  $\alpha$ -lactalbumin and oleic acid (13,14) and the first member in a new family of complexes formed from partially unfolded proteins with fatty acids as integral constituents. HAMLET displays broad anti-tumor activity *in vitro* with a high degree of tumor selectivity (15). Studies in patients and animal models have confirmed this selectivity and demonstrated therapeutic efficacy against several tumor types (16-20). HAMLET has broad tumoricidal activity, unrelated to well-defined cellular responses such as apoptosis (21-23). Essential aspects of the conserved death response to HAMLET have been defined, including the dependence on oncogenic transformation (22), proteasome inhibition (24) and nucleosome-histone binding (25). We have recently identified ion channel activation as a new, unifying mechanism of HAMLET's tumoricidal effect (23). Rapid ion fluxes triggered by HAMLET were shown to initiate changes in morphology, viability, gene expression and MAPK signaling and especially  $\text{Na}^+$  or  $\text{K}^+$  fluxes were essential for these responses to occur.

In a screen for suitable fatty acids cofactors, C18:1, cis-monounsaturated fatty acids were identified as optimal for HAMLET formation (26), suggesting that these fatty acids may share specific structural features required both for HAMLET formation and to engage targets involved in tumor cell death. In contrast, trans-poly/monounsaturated or fully saturated fatty acids failed to form a HAMLET-like complex under near identical experimental conditions (26-29). The contribution of the C18:1, cis fatty acids to the tumoricidal activity of HAMLET has been debated, however. Studies of complexes with high lipid content have recently suggested that unfolded proteins may function solely as "lipid carriers" and that the tumoricidal response is triggered by the LCFAs alone (28,30).

The effect of lipids on host cells is influenced by the protonation state. For example, experiments using anionic inhibitors suggested that the deprotonated form of LCFAs may be the most relevant for cellular uptake (12). The protonation state of oleic acid in HAMLET is unclear due to the phase behavior of oleic acid, which increases the apparent pKa to between 8.0 and 8.5 (31) or 9.85 (32). Oleic acid is expected to be deprotonated in HAMLET, however, as the

complex is formed by ion exchange chromatography at basic pH. Oleate has been used successfully to form HAMLET-like complexes (33), but structural or biological differences between oleic acid and oleate as cofactors in HAMLET have not been examined.

To address this question, we first show by  $^{13}\text{C}$  NMR that oleate and oleic acid produce structurally homologous HAMLET complexes. Using a variety of cellular assays, we subsequently identify dose-dependent effects of oleate on tumor cells, partially overlapping with those of HAMLET. Fatty acids may thus exert some of their essential effects on host cells when in the deprotonated state and when presented in the context of a partially unfolded protein.

## EXPERIMENTAL PROCEDURES

*HAMLET/Oleate-HAMLET production and cell culture*— $\alpha$ -Lactalbumin was purified from defatted human milk by ammonium sulfate precipitation and hydrophobic interaction column and converted to HAMLET by removal of calcium and binding to oleic acid as previously described (14). For Oleate-HAMLET, oleic acid was replaced by sodium oleate (Sigma), dissolved in Tris buffer, pH 8.5, by vortexing.

Lung carcinoma cells (A549) and T-cell lymphoma cells (Jurkat) (ATCC, Manassas, VA) were cultured in RPMI-1640 with non-essential amino acids (1:100), 1 mM sodium pyruvate (all from PAA, Pasching, Austria), 50  $\mu\text{g}/\text{ml}$  Gentamicin (Gibco, Paisley, UK) and 5% fetal calf serum (FCS).

*Circular Dichroism (CD) spectroscopy*—Far- and near-UV CD spectra were collected on HAMLET and Oleate-HAMLET at 25 °C using a Jasco J-810 spectropolarimeter as previously described (14). Lyophilized HAMLET and Oleate-HAMLET were dissolved in phosphate-buffered saline (PBS). Secondary structure predictions were made using K2D3 (34) with 41 input points from 200-240 nm in  $\Delta\epsilon$  unit, calculated as  $\Delta\epsilon = \theta_m/3298$ . Thermal denaturation measurement at 270 nm and 222 nm were collected from 5 to 95 °C at a scan rate of 60 °C  $\text{h}^{-1}$ ; 1 nm wavelength step; 8 s response time. HAMLET and Oleate-HAMLET were reconstituted in PBS at 32 and 43  $\mu\text{M}$  respectively for 270 nm and to 28  $\mu\text{M}$  for 222 nm measurements. Data were presented as above. The extent of unfolding was calculated by taking the ratio of the measured ellipticity at each

temperature to the maximum ellipticity at 270 nm.

*Nuclear magnetic resonance (NMR) spectroscopy*— $^1\text{H}$  and natural abundance  $^{13}\text{C}$  NMR experiments were performed on an 800 MHz (18.8 T) Premium COMPACT 54 mm / UHF DD2 NMR spectrometer with HCN Cold Probe (Agilent Technologies). Lyophilized samples were dissolved in either 100% D<sub>2</sub>O (or 80%/20% H<sub>2</sub>O/D<sub>2</sub>O), 50 mM sodium phosphate buffer, pH 7.4, with 1,4-dioxane added to serve as chemical shift standard (3.75 ppm for  $^1\text{H}$ , 67.3 ppm for  $^{13}\text{C}$ ). Oleic acid and oleate (both from Sigma) were dissolved in 100% methanol for determination of the chemical shifts of protonated and deprotonated carboxyl carbons. The following parameters were used for the  $^{13}\text{C}$  NMR acquisition: temperature 25.0 °C, 90-degree pulse width 14.5 us, acquisition time 0.164 s, spectral width 50000 Hz (-20 to 220 ppm), relaxation delay 9.0 s, data points 16k, and scans 8000.

*Cell death analysis*—A549 lung carcinoma cells ( $5 \times 10^5$  cells/ml in suspension) were treated with HAMLET, Oleate-HAMLET, oleic acid or sodium oleate at concentrations described in Result and cell death was measured by the reduction in ATP level and Trypan Blue exclusion as previously described (23) and Prestoblue cell viability reagent (Invitrogen) according to manufacturer's instructions. Oleic acid was dissolved in ethanol and diluted to 3.5 mM in PBS and sonicated. Sodium oleate was dissolved directly in PBS to 3.5 mM by vortexing and sonication.

*Phase Holographic Imaging*—The HoloMonitor™ M3 digital holographic microscope (Phase Holographic Imaging AB, Lund, Sweden) records 3D information using interfering wave fronts induced by the exposure to a 0.8 mW HeNe laser (633 nm) (35-37). The interference pattern (hologram) is recorded on a digital sensor and is used to reconstruct the amplitude and phase of the object (38,39). 40000 A549 cells were cultured on  $\mu$ -Slide I coated with ibiTreat (ibidi, Martinsried, Germany) overnight. The cells were washed and replaced with fresh serum-free RPMI medium and exposed to 35 mM HAMLET, 175 mM oleate or oleic acid for 3 hours at 37 °C, 5% CO<sub>2</sub>. 5% FCS was added after 1 h of incubation. The holograms were captured with an imaging time of 2.4 msec, every 15-minute interval for the first hour and at 90, 120 and 180 minutes after treatment.

*Intracellular ion concentrations and ion fluxes*—

The relative, free intracellular concentrations of  $\text{Ca}^{2+}$  ( $[\text{Ca}^{2+}]_i$ ) and  $\text{Na}^+$  ( $[\text{Na}^+]_i$ ) were estimated using the calcium indicator Fluo4 NW and the sodium fluorophore CoroNa Green, respectively (Invitrogen), as previously described (23). For estimation of  $\text{K}^+$  fluxes, the FluxOR™ potassium ion channel assay (Invitrogen) was used according to the manufacturer's instructions. Briefly, cells were incubated with FluxOR™, which is a  $\text{Ti}^+$  indicator. An increase in fluorescence signal, which was measured at 535 nm after excitation at 485 nm, corresponds to an influx of  $\text{Ti}^+$ , indicating opening of potassium channels.

*Confocal microscopy*—A549 lung carcinoma cells were seeded overnight at 37 °C, 5% CO<sub>2</sub> on 8-well chamber slides (Nalge Nunc, Rochester, NY). The cells were washed twice with serum-free RPMI medium and incubated with HAMLET, Oleate-HAMLET, oleic acid or sodium oleate at equimolar concentration for a period of 30 minutes to 3 hours in RPMI medium at 37 °C, 5% CO<sub>2</sub>. 5% FCS were added after 1 h of incubation. The cells were fixed with 3.7 % formaldehyde (FA). The supernatant with detached cells were fixed with FA separately and cytopinned at 500 rpm for 5 min onto L-lysine coated glass slides (Thermo Scientific) for analysis. The cells were stained with 100  $\mu\text{l}$  Nile Red (9-diethylamino-5H-benzo[ $\alpha$ ]phenoxazin-5-one, 100  $\mu\text{g}/\text{ml}$ ) (Sigma-Aldrich, St. Louis, MO, USA) for 5 min at room temperature. Nuclei were stained using DRAQ5 (eBioscience, San Diego, CA).

The slides were analyzed with LSM 510 META system (Carl Zeiss, Jena, Germany) with two spectra settings: Excitation with 488 nm and emission analysis with a Band Pass 505-550 filter, for yellow-gold fluorescence; Excitation with 543 nm and emission analysis with a Long Pass 585 filter, for red fluorescence

*Transcriptomics*—A549 cells (200 000/well) were allowed to adhere overnight on a 6-well plate (TPP, Trasadingen, Switzerland). After exposure to HAMLET (1 hour, 35  $\mu\text{M}$ ), oleic acid or sodium oleate (both from Sigma-Aldrich, St. Louis, MO, USA, 1 hour, 175  $\mu\text{M}$ ), RNA was extracted (RNeasy Mini Kit, QIAGEN). The samples were analyzed using standard Affymetrix protocols. The raw data was normalized using RMA (40) as provided by R and Bioconductor (<http://www.r-project.org>). Probe sets with a p value < 0.05 and log2 fold

change > 1 were considered differentially expressed. To functionally characterize the resulting gene lists, Database for Annotation, Visualization and Integrated Discovery (DAVID) (41) and Ingenuity Pathway Analysis (IPA) (Ingenuity Systems®) were employed.

**Real-Time PCR**—Total RNA was reverse-transcribed using SuperScript™ III First-Strand Synthesis System for RT-PCR (Invitrogen) and quantified in real time using QuantiTect SYBR Green based PCR Kits for mRNA (Qiagen) on a Rotor Gene Q (Qiagen). qRT-PCR reactions were run in biological duplicates and triplicates and gene expression was done based on the comparison with GAPDH expression.

**Metabolomics**—HAMLET-treated cells were washed with phosphate-buffered saline, followed by the addition of ice-cold methanol to stop metabolic activity. Subsequently, cells were scraped into the solvent and transferred to a reaction tube. Six replicates consisting of three pooled wells each were collected per sample group. For extraction, the methanol phase was removed from the cells, dried and stored at -80°C. Cell pellets were dried in a FreeZone 2.5 lyophilizer (Labconco, Kansas City, MO, USA), homogenized using a Mini-Beadbeater (BioSpec Products, Bartlesville, OK, USA) and extracted a pre-cooled methanol-isopropanol-water (3:3:2) mixture. A mixture of internal retention index markers was prepared using fatty acid methyl esters of C8, C9, C10, C12, C14, C16, C18, C20, C22, C24, C26, C28 and C30 linear chain length, dissolved in chloroform at a concentration of 0.8 mg/ml (C8-C16) and 0.4 mg/ml (C18-C30). 1 ml of this RI mixture was added to the dried extracts. 10 µl of a solution of 40 mg/ml of 98% pure methoxyamine hydrochloride (CAS No. 593-56-6, Sigma, St. Louis MO) in pyridine (silylation grade, Pierce, Rockford IL) was added and shaken at 30 °C for 90 min to protect aldehyde and ketone groups. 90 µl of MSTFA. 1%TMCS (1 ml bottles, Pierce, Rockford IL) was added for trimethylsilylation of acidic protons and shaken at 37 °C for 30 min. A Gerstel (Mülheim an der Ruhr, Germany) automatic liner exchange system with multipurpose sample MPS2 dual rail was used to inject 0.5 ml sample to a Gerstel CIS cold injection system (Gerstel). Samples are injected into a 50 °C injector port and held for 1 minute. They are then separated using an Agilent 6890 gas chromatograph, equipped with a 30 m long, 0.25 mm i.d. Rtx5Sil-MS column (Restek (Bellefonte, PA, USA), 0.25mm 5% diphenyl

film and additional 10 m integrated guard column) by ramping to 330 °C at 20 °C min<sup>-1</sup>, and holding for 5 min. Mass spectrometry was performed on a Leco Pegasus IV time-of-flight mass spectrometer (St Joseph, MI, USA) with a 280 °C transfer line temperature, -70 eV electron ionization and an ion source temperature of 250 °C. Mass spectra were acquired from m/z 85 to 500 at 17 spectra/second and 1850 V detector voltage. Result files were processed using the metabolomics BinBase database. All database entries in BinBase were matched against the Fiehn mass spectral library of B1200 authentic metabolite spectra using retention index and mass spectrum information or the NIST05 commercial library (<http://fiehnlab.ucdavis.edu/Metabolite-Library-2007/>). Identified metabolites were included if present within at least 50% of the samples of each treatment group (as defined in the SetupX database).

## RESULTS

**Oleate is the functional cofactor in HAMLET**—The protonation state of bound oleic acid in HAMLET was determined by natural abundance <sup>13</sup>C NMR spectroscopy. Protonated and deprotonated carboxyl groups were clearly distinguishable from the overlay of the two spectra (Fig. S1A). Carboxyl carbon of oleate showed a chemical shift at 182 ppm whereas that of oleic acid was 177 ppm. Both sodium oleate and oleic acid dissolved in methanol or sodium phosphate buffer gave rise to a 130 ppm peak, which corresponds to the olefinic carbons. Peaks further upfield correspond to the aliphatic carbons. The natural abundance of <sup>13</sup>C in HAMLET was determined (Fig. 1). HAMLET showed a prominent peak at 182 ppm and a broad 130 ppm peak, a clear difference from free fatty acid. A minor bound oleic acid peak was also detected. An additional peak at 175 ppm, verified to be the carboxyl carbon of residual EDTA in the conversion process was also recorded. These results identify oleate as the lipid cofactor in the HAMLET complex.

**Oleate and oleic acid form homologous HAMLET complexes**—To confirm the protonation state of oleic acid in HAMLET, ion exchange matrices were pre-conditioned with oleic acid or sodium oleate, as described (14). The preconditioned columns were loaded with EDTA-treated, partially unfolded human α-lactalbumin and eluted using a NaCl gradient.

HAMLET and Oleate-HAMLET eluted as sharp peaks at 0.7 M NaCl (Fig. S1A and B). By near-UV CD spectroscopy (250-320 nm wavelength) (Fig. S1C), HAMLET and Oleate-HAMLET showed a similar decrease in signal intensity at 270 nm as compared to native human  $\alpha$ -lactalbumin (14). By far-UV CD spectroscopy, both complexes were shown to retain similar secondary structure contents (Fig. S1D) (14). Further structural characterization by thermal denaturation, followed by CD spectroscopy confirmed the structural similarity (Fig. S1E-G) but unfolding of HAMLET occurred at a lower temperature. The melting temperature,  $T_m$ , estimated from the thermal denaturation profiles for native  $\alpha$ -lactalbumin and HAMLET were  $\sim 60^\circ\text{C}$  and  $\sim 31^\circ\text{C}$ , respectively (42,43) and the recorded  $T_m$  Oleate-HAMLET was  $\sim 35^\circ\text{C}$  (Fig. S2E). Natural abundance  $^{13}\text{C}$  NMR of Oleate-HAMLET confirmed that oleate is the bound state in both complexes, showing a prominent peak at 182 ppm and a broad 130 ppm peak (Fig. 1).

HAMLET and Oleate-HAMLET thus show a high degree of structural similarity, including almost identical elution profiles, similar secondary and tertiary structure characteristics and melting temperature.

*Tumor cell death in response to HAMLET, Oleate-HAMLET, oleate and OA-* To ensure that Oleate-HAMLET reproduces the tumoricidal effect of HAMLET cell death was quantified in A549 lung carcinoma cells exposed to the respective complex (Fig. 2A). No difference in dose-dependent cytotoxicity was recorded between HAMLET and Oleate-HAMLET ( $>80\%$  dead cells after 3 hours at  $35\ \mu\text{M}$ ), (Fig. S2A).

The lipid concentration in the HAMLET/Oleate-HAMLET complexes was determined as approximately 1:4 or 1:5 by acid hydrolysis and GC/MS (26). At a concentration corresponding to five oleate molecules ( $175\ \mu\text{M}$ ) per protein ( $35\ \mu\text{M}$   $\alpha$ -lactalbumin), oleate or oleic acid did not alter the viability of lung carcinoma cells (3 hours) compared to control (n.s., Fig. S2B). At higher lipid concentrations (15 times), oleate was more efficient as a cytotoxic agent than oleic acid (Fig. S3B). At 25 times, all cells underwent rapid lysis, confirming well-known effects of high lipid concentrations on cellular integrity.

Effects on tumor cell morphology were compared by confocal microscopy (Fig. S2C). In

response to HAMLET and Oleate-HAMLET ( $35\ \mu\text{M}$ , 1 hour) the cells rounded up and detached. By real time holography imaging (Fig. 2B), HAMLET and Oleate-HAMLET caused a parallel reduction in cell surface area and an increase in height. Less pronounced morphological changes were observed in cells treated with oleate alone and oleic acid left the cells largely unchanged. By transmission light microscopy, all treated cells showed an increase in granularity indicative of lipid droplet formation, suggesting uptake of both the protonated and deprotonated fatty acid (Fig. S2C).

*Differences in ion channel activation by HAMLET and oleate/oleic acid-HAMLET* triggers ion fluxes across cell membranes and ion channel inhibitors, blocking such fluxes prevent HAMLET uptake as well as tumor cell death (23). To address if oleate is an independent trigger of ion fluxes, ion channel activation was quantified in lung carcinoma or Jurkat cells preloaded with  $\text{Ca}^{2+}$  or  $\text{Na}^{+}$  fluorophores or thallium, a surrogate marker for  $\text{K}^{+}$ . HAMLET and Oleate-HAMLET triggered rapid  $\text{Na}^{+}$ ,  $\text{K}^{+}$  and  $\text{Ca}^{2+}$  fluxes of similar magnitude (Fig. 2C). A weak  $\text{Na}^{+}$  and  $\text{K}^{+}$  response to oleate was recorded but no  $\text{Ca}^{2+}$  response was detected. Increasing the oleate concentration to a cytotoxic level of  $500\ \mu\text{M}$  did not entirely reproduce the ion flux patterns induced by HAMLET (Fig. S2D).

*The metabolic response to oleate-HAMLET* causes metabolic paralysis in tumor cells, with a reduction  $>60\%$  in abundance of metabolites after one hour (22). To examine the effects of oleate on the metabolic response, oleate-treated cells (60 minutes,  $175\ \mu\text{M}$ ) were subjected to a non-targeted metabolite profiling by gas chromatography-mass spectrometry (GC-MS), (Fig. 3A). After HAMLET treatment, 34 of the 49 altered metabolites were reduced compared to 16 of 24 in oleate-treated cells (Fig. 3B). An increase in oleic acid, elaidic acid and icosenic acid was detected, possibly reflecting an increase in fatty acid uptake by the cells. Decreased metabolites after oleate-treatment included pyrophosphotase, aconitic acid, glycine, taurine and TRIS (Fig. S3). In contrast, only three metabolites were altered by oleic acid (60 minutes,  $175\ \mu\text{M}$ ), with an increase in pentadecanoic acid and tyrosine (3.6- and 3.8-fold) and decrease in elaidic acid compared to

control (4-fold) (Fig. S3). The results suggested major differences in the effects of HAMLET and oleate on tumor cell metabolism.

To further characterize the metabolic response, HAMLET- or oleate- treated cells were subjected to targeted metabolite profiling of fatty acids, amino acids and citric acid cycle constituents (Fig. 3C). In total, 43 metabolites were significantly altered by HAMLET treatment ( $p$  value  $< 0.05$ , Supplementary Table S1). Consistent with the non-targeted profiling, an increase in fatty acid metabolites was detected (stearic-, palmitic-, methylhexadecanoic-, elaidic-, azelaic-, arachidic- and myristic acid, Fig. 3C). There was a loss of early metabolites in the citric acid cycle (citric-, isocitric- and alpha-ketoglutaric acid) and an accumulation of late metabolites (succinic-, fumaric- and malic acid). Furthermore, metabolites feeding into the citric acid cycle were reduced, in particular, sorbitol, fructose and glycerol-alpha-phosphate and amino acids (asparagine, isoleucine, phenylalanine, serine, threonine and valine). In parallel, a late glycolytic intermediate (3-phosphoglycerate) and a carbohydrate metabolism derivative (5-hydroxy-methyl-2-furoic acid) were increased after HAMLET treatment.

In the targeted metabolic profiling, the effects of oleate were at least as pronounced as for HAMLET. In total, 61 metabolites were significantly altered ( $p$  value  $< 0.05$ , Supplementary Table S1). Fatty acid metabolites were less strongly affected than by HAMLET (azelaic-, elaidic- and stearic acid were increased) and the loss of early and accumulation of late metabolites in the citric acid cycle was similar to HAMLET. The deregulation of amino acid metabolism was more apparent in oleate- than in HAMLET treated cells, with an additional nine decreased amino acids or amino acid derivatives.

The results suggest that HAMLET and oleate have similar effects on the TCA cycle but that the pronounced effects of HAMLET on overall tumor cell metabolism are not reproduced by oleate (175  $\mu$ M). The general decrease in amino acids in HAMLET and oleate treated cells might indicate that amino acids are used as an alternative source of energy when the proximal TCA cycle is blocked.

*Effects of HAMLET and oleate/oleic acid on gene expression*-Genome wide transcriptomic analysis was used to further compare the cellular effects of HAMLET and oleate. RNA samples from lung carcinoma cells treated with HAMLET (35  $\mu$ M) or oleic acid or oleate (175  $\mu$ M) were hybridized to Affymetrix Whole Genome Microarrays U219. The resulting hybridization profiles were assessed pre- and post-RNA normalization. For statistical analysis normalized data were linear model fitted and  $p$ -values were estimated using an empirical Bayes approach. Genes with a log2-fold change  $> 1$  and  $p$ -value  $< 0.05$  were considered differentially expressed. The transcriptional response to oleic acid was restricted to two genes with a log2-foldchange  $> 1$  and  $p$ -value  $< 0.05$ , EGR1 and PKD4.

A heatmap of the top 200 differentially expressed genes by fold change is shown in Figure 4A. A pronounced effect of HAMLET on gene expression was observed (74 upregulated and 128 suppressed genes) compared to oleate (19 upregulated and 2 suppressed genes), 19 of which overlapped with HAMLET (Fig. 4B-C). The suppressed genes (Fig. 4B) include genes associated with DNA damage and repair and cell cycle regulation (enrichment scores of 2.37 and 1.8, respectively). Increased transcription of genes associated with cell death and transcriptional regulation was recorded (enrichment scores of 4.99 and 4.68, respectively).

Top canonical pathways affected by HAMLET included the MAPK signaling pathway, cytokine-cytokine receptor interactions, chemokine- and NOD-like receptor signaling pathway, as well as genes in "pathways in cancer" (Fig. 4D). Four p38 regulators were upregulated including HSPA6 (2.8-fold) and three dual-specificity phosphatases (DUSP1, DUSP5 and DUSP6, 2.4, 2.1 and 1.1-fold, respectively) acting as feedback regulators of MAPK signaling. Transcription factors that respond to DNA damage and ER stress were upregulated (GADD45A/B and DDIT3/4, 1.5, 1.8, 1.8 and 1.9-fold) as were genes implicated in cell death, including ATF3, MAP4K5 and BIRC3. Transcription factor analysis identified 37 NFkB regulated transcripts (z-score 4.72 for NFkB) including FOS, ATF3, IL8, EGR1, CCL20 and JUN (all with a log2 foldchange  $> 4$ ). The 27 p53-dependent transcripts (z-score 2.85 for p53) included PHLDA1, GADD45A, DDIT3 and DDIT4. Ten HDAC-regulated transcripts

were suppressed (Z-score -2.94, for Hdac), consistent with previously published effects of HAMLET on histone acetylation (44).

Networks associated with cancer, cellular movement and morphology, cell cycle, cellular assembly and organization were either exclusively found in the HAMLET treated cells or had a lower score in the oleate (175  $\mu$ M) sample (Fig. 4F). Pathways overlapping between HAMLET and oleate included cytokine-cytokine receptor interactions, MAPK-, chemokine-, and NOD-like receptor signaling pathway, although the signals were weaker in oleate treated cells (Fig. 4D).

The 183 HAMLET specific genes were associated with transcription (39 genes), intracellular signaling cascades (25 genes), phosphate metabolic processes (20 genes) and cell cycle (21 genes). Ingenuity Pathway Analysis (IPA) identified 131 cancer-related genes, half of which formed a cancer-related network including NFKB1A, MAP3K8, SMAD7 and DUSP6 (Fig. 4G). Two oleate specific genes were CUL3, involved in polyubiquitination and endosome maturation, and ANGPTL4, which acts as a regulator of angiogenesis.

Differential gene expressions of FOS, JUN, TNFRSF10D and DUSP1 for oleate (175  $\mu$ M) were confirmed by RT-PCR (Figure 4E). Two additional genes that were not found to be differentially expressed in the microarray, HSPA6 and DDX3X, were also included and validated as their expression levels were less than the predetermined cut-off expression value.

The results suggest that most HAMLET effects on gene expression and signaling are not reproduced by oleate. Minor overlap between oleate and HAMLET induced responses was observed.

*Transcriptional response to higher oleate concentrations*-To address if a higher lipid concentration might reproduce the effects of HAMLET more fully, tumor cells were treated with 500  $\mu$ M of oleate. A heatmap of significantly regulated genes is shown in Fig. 5A. The higher oleate concentration triggered a significant response (157 upregulated and 158 suppressed genes) showing more overlap with the HAMLET sample (107/202 genes, Fig. 5B-C). Overlapping pathways included MAPK signaling-, cytokine- and chemokine signaling pathways (Fig. 5D). A significant overlap was also seen in broad cancer related networks,

where the high oleate sample stimulated a larger number of genes than HAMLET.

By DAVID, genes specific for the high oleate sample were shown to be associated with RNA processing (16 genes), intracellular transport (13), cellular macromolecule catabolic processes (15), cell cycle regulation (18) and RNA splicing (12). By IPA, 54 oleate specific, cancer-related genes were identified, half of which formed a cancer-related network including MYC, KRAS, PXN and MET (Fig. 5G). After exclusion of genes shared with the high oleate samples, genes that remained unique to HAMLET were mainly involved in intracellular signaling cascades (15 genes), regulation of biosynthetic process (12) and phosphate metabolic processes (12 genes). Networks involved in the maintenance or cell death and survival were also regulated.

The results suggest that important effects on gene expression are shared between HAMLET and oleate and that the extent of overlap between HAMLET and oleate-induced genes increased with the oleate concentration. Even at high oleate concentrations, the majority of oleate-induced genes were not regulated in HAMLET treated cells, however.

## DISCUSSION

Oleic acid is a key constituent of the HAMLET complex, but the extent to which the lipid contributes to tumor cell death has remained unclear, as have the mechanisms involved. HAMLET is optimally formed from 18-carbon, monounsaturated, cis-fatty acids and partially unfolded  $\alpha$ -lactalbumin, in contrast to unsaturated trans- or saturated fatty acids, which fail to form complexes with the protein. In the present study, we show that the deprotonated form of oleic acid (oleate) is an active lipid cofactor in HAMLET formation and that when presented in the context of partially unfolded  $\alpha$ -lactalbumin, oleate acts directly on the cell death machinery, through membrane sensing, uptake and metabolic processing. The lipid *per se* did not reproduce the cell death response to HAMLET or the metabolic paralysis, however confirming that the protein and lipid are both required for the full-fledged tumoricidal response to HAMLET. At higher oleate concentrations, a larger part of the HAMLET profile was reproduced, but major differences in cellular response profiles were still present. Oleate recognition by tumor cells is thus



quantitatively and qualitatively altered in the context of partially unfolded  $\alpha$ -lactalbumin.

A number of HAMLET-related complexes have been prepared using different  $\alpha$ -lactalbumin variants (45,46), aggregation states (27), related proteins like equine lysozyme (47), bovine  $\beta$ -lactoglobulin (bLG) or the  $\alpha$  isoform of pike parvalbumin (28). Some of these complexes, which contain a high number of oleic acid molecules (27), have been proposed to permeabilize cells by virtue of their fatty acid content alone, giving rise to suggestions that the protein acts merely as a fatty acid delivery vehicle (48-50). It may be speculated that upon interaction of tumor cell membranes with several HAMLET molecules, membrane patches with higher lipid concentrations might be formed, resulting in membrane disruption and necrosis. The present study clearly demonstrates that HAMLET and Oleate-HAMLET complexes carry a limited number of lipid moieties and that distinct cellular responses including ion fluxes and specific death pathways are activated, arguing against non-specific membrane disruptions as a mechanism of tumor cell death. While we show that the lipid is active in the context of the partially unfolded protein, we found no evidence of independent cytotoxic activity, at concentrations equimolar to those present in HAMLET. At higher concentrations, the lipid was cytotoxic on its own, however. Importantly, cellular response profiles at either lipid concentration were not those involved in necrosis, as would have been expected if HAMLET merely permeabilizes the cell membrane due to its lipid content.

In this study, HAMLET and oleate were shown to trigger overlapping but also fundamentally different responses in tumor cells. At equimolar concentrations, oleate alone was shown to activate  $\text{Na}^+$  fluxes but not the full ion channel repertoire seen in HAMLET treated cells. This limited effect may explain the low transcriptional response to oleate (175  $\mu\text{M}$ ) and its lack of cytotoxicity. At higher oleate concentrations, effects on ion fluxes were stronger, compatible with the transcriptional changes and the cytotoxic response and overlapping effects on transcription included cancer related pathways, MAPKs genes and innate immune response genes. Ion fluxes were not identical to HAMLET, however, as oleate (500  $\mu\text{M}$ ) caused partial activation of  $\text{Na}^+$  and  $\text{Ca}^{2+}$ , compatible with the differences in transcriptomic profiles. The results emphasize

that the HAMLET complex is functionally defined, in part by oleate but possesses unique additional properties not reproduced by the lipid.

Cancer metabolism is optimized for cell growth and survival under conditions of metabolic stress (51). In addition to the shift in glycolysis known as the Warburg effect, there is evidence that fatty acid oxidation contributes to the metabolic changes accompanying oncogene over-expression (52,53). We detected two major alterations in cellular metabolism, one entailing a global metabolic paralysis, which was seen exclusively in HAMLET treated cells and the second a shift in lipid metabolites, citric acid cycle constituents and amino acids, seen both in HAMLET and oleate-treated cells but not in oleic acid treated cells. HAMLET and oleate both increased fatty acid metabolism, consistent with lipid recognition and uptake by tumor cells, but oleic acid had no significant effect. The inertia of cells to the protonated acid was surprising but may indicate that the deprotonated fatty acid is the more active cellular agonist, as indicated by previous, cellular uptake studies (54).

Critical starting metabolites in the citric acid cycle were less abundant after HAMLET treatment, consistent with a loss of pyruvate and inhibition of glycolysis. Previously, HK1 and the glycolytic machinery have been shown to influence HAMLET sensitivity and direct binding of HAMLET to the kinase has been detected (22). In parallel, an increased rate of amino acid catabolism and a buildup of metabolites towards the end of the cycle was observed, further suggesting that HAMLET may be forcing tumor cells to use glucose-independent metabolic pathways and amino acid catabolism as an energy source potentially compensating for the inhibition of glycolysis. Oleate alone seemed to direct metabolism towards amino acid catabolism, suggesting that the deprotonated lipid is deregulating the TCA cycle, but not triggering death. Amino acid metabolism is important for cancer cell survival and the use of amino acid metabolites for energy production has been proposed to offer a glucose-independent solution to ATP generation in tumor cells (55,56).

This study resolves, for the first time, the question of the lipid contribution to the tumoricidal activity of HAMLET. The results confirm that the lipid is a necessary constituent to achieve the tumoricidal effect of the complex (14). This was previously inferred from the

finding that partially unfolded  $\alpha$ -lactalbumin protein alone does not trigger tumor cell death (57,58). On the other hand, oleic acid alone was largely inactive, presenting somewhat of a paradox. In this study, the deprotonated lipid is shown to be a fully functional cellular agonist both alone and in the context of  $\alpha$ -lactalbumin. The oleate effect was concentration-dependent, as higher oleate concentrations reproduced more of the cellular response to HAMLET. Important

qualitative differences were observed between the HAMLET complex and oleate alone at equimolar or higher concentrations, however. While effects on gene expression were overlapping, significant sets of genes were uniquely responsive to oleate or HAMLET, respectively. We conclude that partially unfolded  $\alpha$ -lactalbumin is a suitable partner for oleate, to offer tumor cells the “kiss of death”.

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## FOOTNOTES

This study was supported by the Sharon D. Lund foundation grant and the American Cancer Society, the National Institutes of Health, National Cancer Institute grant U54 CA 112970, the Swedish Cancer Society, the Medical Faculty (Lund University), the Söderberg Foundation, the Segerfalk Foundation, the Anna-Lisa and Sven-Erik Lundgren Foundation for Medical Research, the Knut and Alice Wallenberg Foundation, the Lund City Jubileumsfond, the John and Augusta Persson Foundation for Medical Research, the Maggie Stephens Foundation, the Gunnar Nilsson Cancer Foundation, the Inga-Britt and Arne Lundberg Foundation, the HJ Forssman Foundation for Medical Research and the Royal Physiographic Society. Support was also obtained from the Danish Council for Independent Research (Medical sciences).

## FIGURE LEGENDS

**FIGURE 1. Natural abundance  $^{13}\text{C}$  NMR spectra of (A) HAMLET and (B) Oleate-HAMLET.** Carboxyl and olefinic carbon regions of  $^{13}\text{C}$  NMR, either in 100% D<sub>2</sub>O (or 80%/20% H<sub>2</sub>O/D<sub>2</sub>O), 50 mM sodium phosphate buffer, pH 7.4. Chemical shift positions of bound oleate/oleic acid, residual EDTA and olefinic carbons are marked. HAMLET and Oleate-HAMLET showed an identical prominent peak at 182 ppm and a broad 130 ppm peak, indicating that oleate is bound to both complexes.

**FIGURE 2. HAMLET and Oleate-HAMLET trigger tumor cell death, morphological change and ion fluxes.** (A) Cytotoxic responses in A549 lung carcinoma cells treated with HAMLET or Oleate-HAMLET (35  $\mu\text{M}$ ), quantified by Trypan Blue exclusion and ATP levels. Sodium oleate- or Oleic acid (175  $\mu\text{M}$ ) were not active at the molar concentration present in HAMLET/Oleate-HAMLET. (B) Morphological response to HAMLET, Oleate-HAMLET (1 hour, 35  $\mu\text{M}$ ), Sodium-oleate or Oleic acid (1 hour, 175  $\mu\text{M}$ ), recorded by holographic imaging. Rounding up of cells was observed after HAMLET and Oleate-HAMLET treatment and Oleate caused a partial effect. (C) The relative free, intracellular concentrations of  $\text{Na}^+$ ,  $\text{K}^+$ , or  $\text{Ca}^{2+}$  ( $[\text{Na}^+]_i$ ,  $[\text{K}^+]_i$ , and  $[\text{Ca}^{2+}]_i$ ) were estimated in A549 cells ( $\text{K}^+$  and  $\text{Ca}^{2+}$ ) or Jurkat cells ( $\text{Na}^+$ ) by fluorescence spectrometry using CoroNa Green, FluxOR, or Fluo-4. Compared to HAMLET (35  $\mu\text{M}$ ), oleate (175  $\mu\text{M}$ ) caused a moderate efflux of potassium and no influx of calcium or sodium in A549 or Jurkat cells, respectively.

**FIGURE 3. Metabolic responses to HAMLET, oleate or oleic acid.** Metabolites were quantified by GC/MS in extracts of A549 cells treated with HAMLET (35  $\mu\text{M}$ ) or oleate (175  $\mu\text{M}$ ) for 60 min. (A) Heatmap of differentially abundant metabolites. A global decrease in metabolite abundance was detected in the HAMLET samples. (B) Frequency distribution of log 2 transformed fold changes in signal intensity between HAMLET treated and oleate treated cells. (C) Targeted metabolite profiling. Change in abundance of lipids, amino acids and the citric acid cycle metabolites in HAMLET- or oleate treated cells.

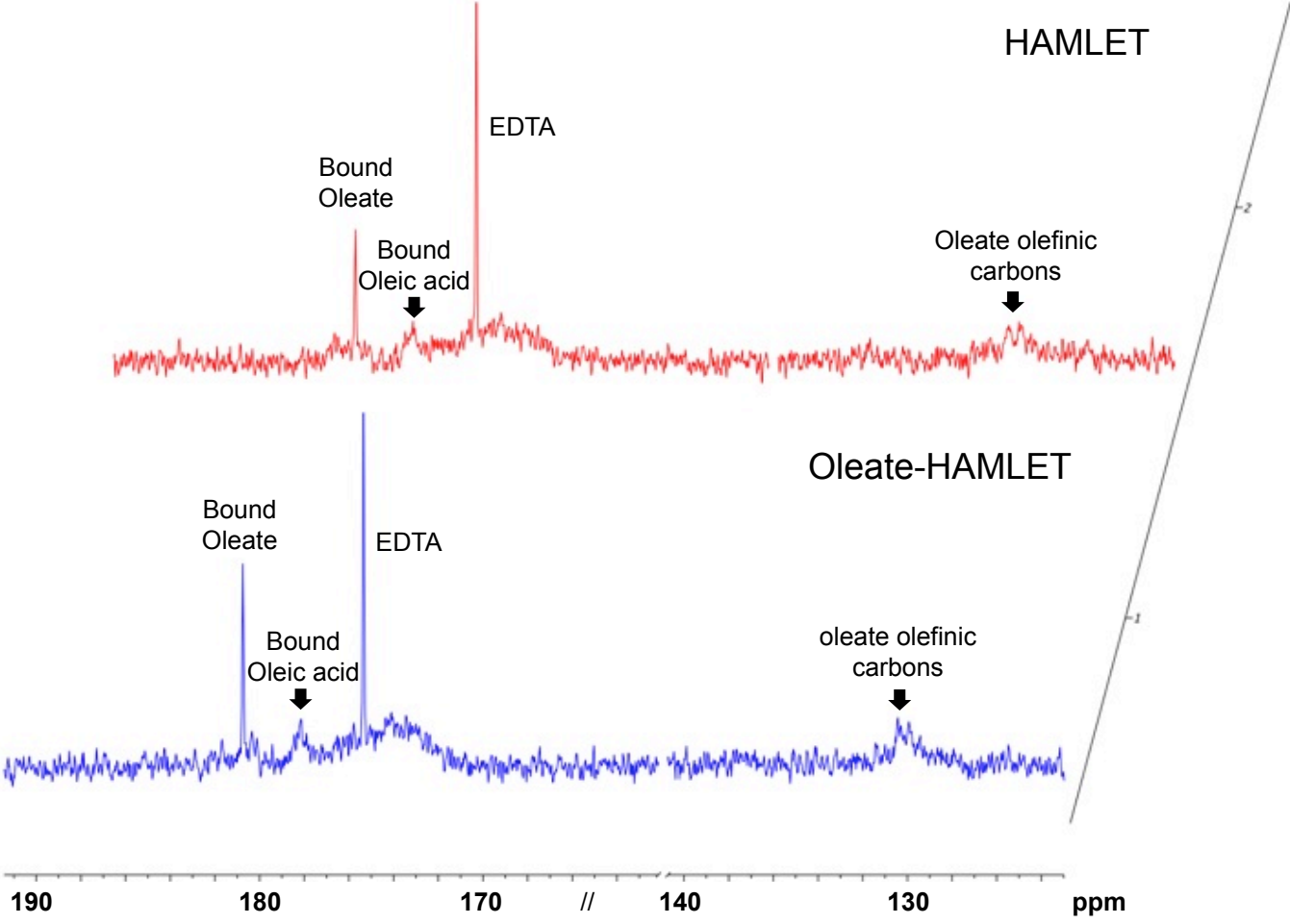
**FIGURE 4. Effects of HAMLET or oleate on gene expression.** A 549 lung carcinoma cells were treated with HAMLET (35  $\mu\text{M}$ ) or oleate (175  $\mu\text{M}$ ) and RNA was harvested after 1 hour. (A) Heatmap of top 200 differentially expressed genes ( $\log_2\text{-foldchange} > 1$  and  $p\text{-value} < 0.05$ ) by fold change. (B) Up- and down-regulated genes in HAMLET- versus oleate-treated samples. More genes were differentially regulated by HAMLET than by oleate. (C) HAMLET caused differential expression of 202 genes, 19 of which were shared by oleate. (D) Shared or specific canonical pathways regulated by HAMLET or oleate. (E) Confirmation of gene expressions for FOS, JUN, HSPA6, DDX3X, TNFRSF10D and

DUSP1 using RT-PCR (F) A network of cancer-related genes differentially regulated by HAMLET. Downregulated genes were colored in green and upregulated genes in red. Expression values from the oleate sample were overlaid onto the same network, showing that most genes were not differentially regulated in the oleate sample.

**FIGURE 5. Gene expression in response to HAMLET or oleate (500  $\mu$ M).** A 549 lung carcinoma cells were treated with HAMLET (35  $\mu$ M) or oleate (500  $\mu$ M) and RNA was harvested after 1 hour. (A) Heatmap of top 200 differentially expressed genes ( $\log_2$ -foldchange  $> 1$  and p-value  $< 0.05$ ). (B) Up- and down-regulated genes for HAMLET and oleate 500 samples. More genes were differentially regulated for oleate 500 compared to HAMLET. (C) Venn diagram showed unique and overlapping genes in HAMLET- or oleate 500 samples. (D) Shared or specific canonical pathways regulated by HAMLET or oleate. (E) Top genes unique to HAMLET and oleate 500 were shown. (F) A network of cancer-related genes differentially regulated by oleate 500. Expression values from the oleate sample were overlaid onto the same network, showing that most genes were not differentially regulated in the HAMLET sample. Downregulated genes were colored in green and upregulated genes in red.



Fig. 1



**Fig. 2**

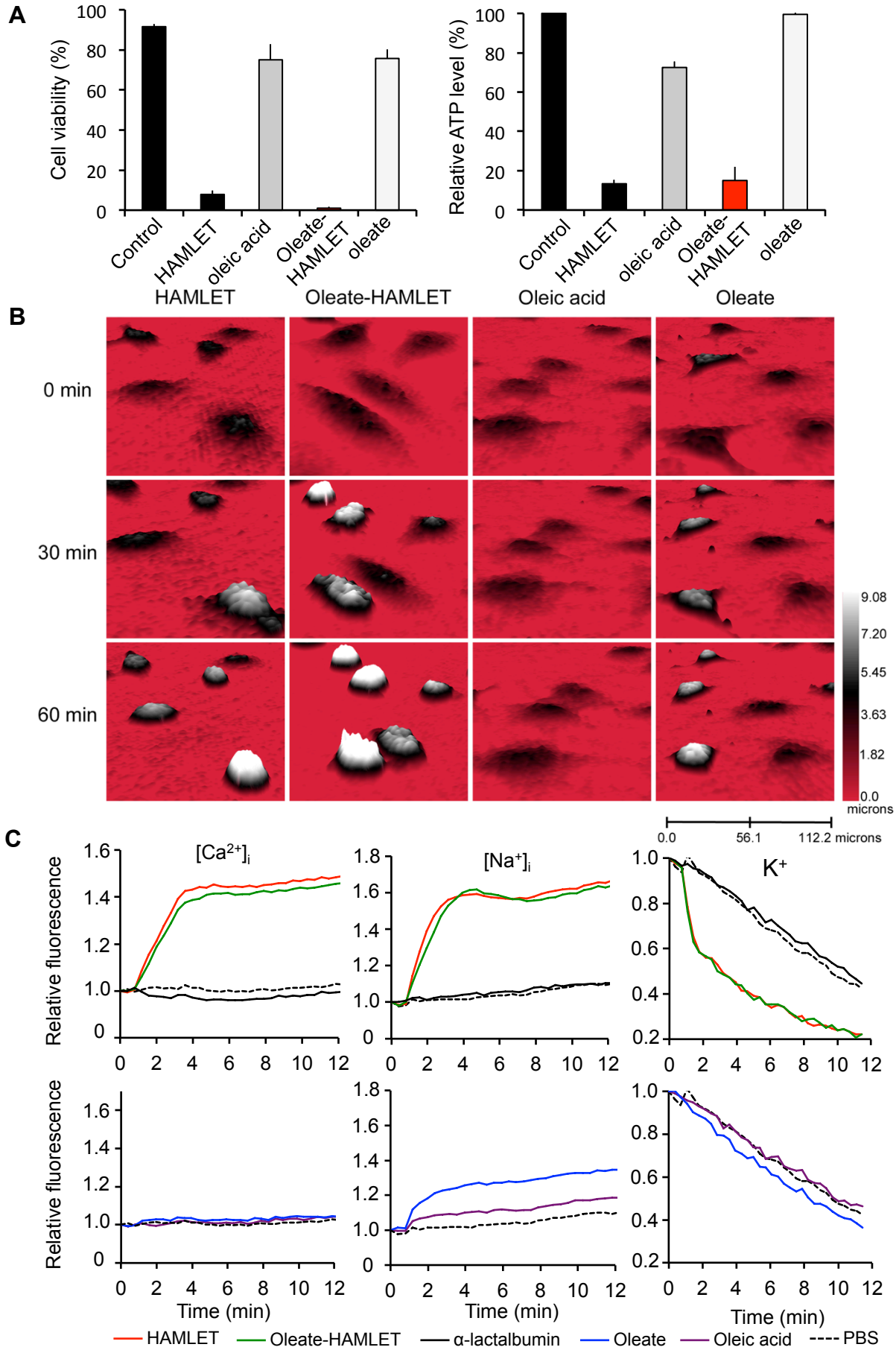
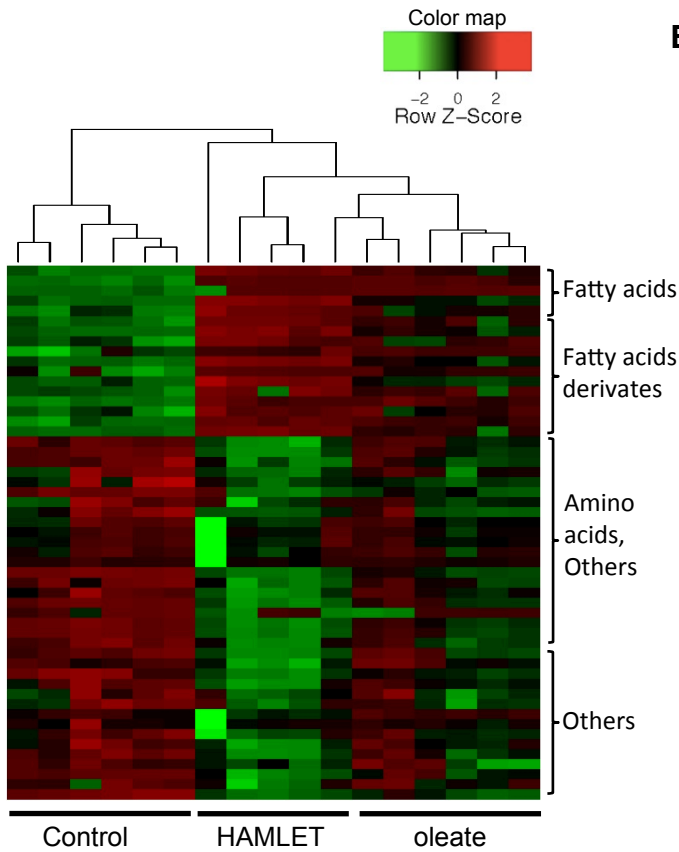
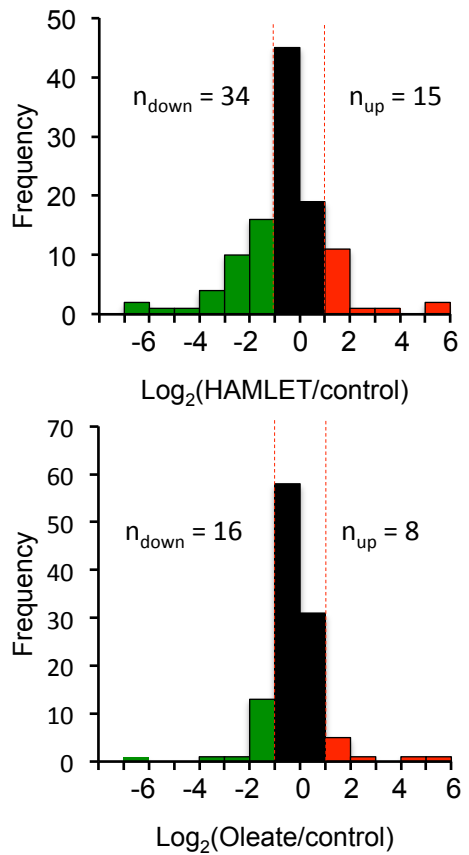


Fig. 3

A



B

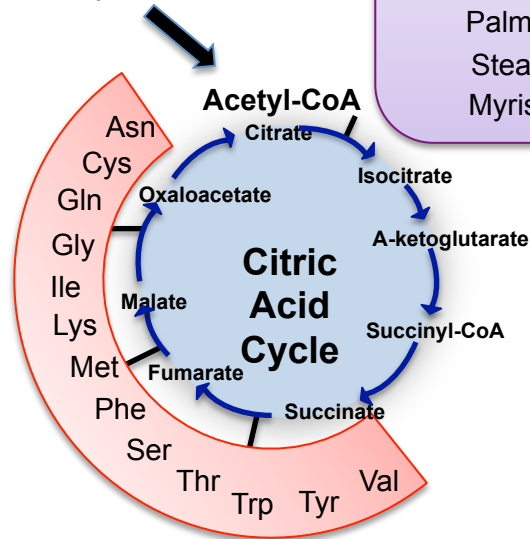


C

Amino Acid Metabolism

	H	O
Asn	-1.56	-1.67
Cys	n.s.	-1.77
Glu	n.s.	-1.31
Gln	n.s.	-1.65
Gly	n.s.	-1.59
Ile	-1.53	-1.60
Leu	n.s.	-1.40
Lys	n.s.	-1.49
Met	n.s.	-1.54
Phe	-1.54	-1.74
Ser	-1.43	-1.60
Thr	-1.39	-1.69
Trp	n.s.	-1.57
Tyr	n.s.	-1.64
Val	-1.46	-1.64

Glucose  
↓ HK1  
Glucose-6-phosphate  
↓  
3-phosphoglycerate  
↓  
Pyruvate



Lipid Metabolism

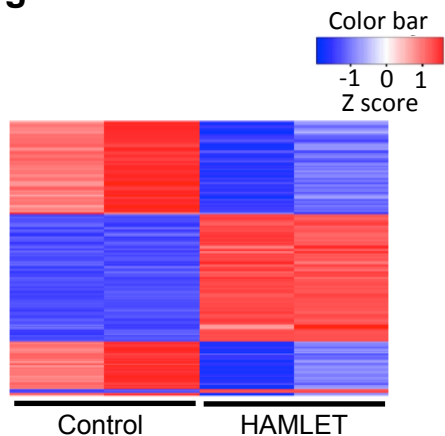
Arachidic acid  
Azelaic acid  
Elaidic acid  
Icosenoic acid  
Methylhexadecanoic acid  
Palmitic acid  
Stearic acid  
Myristic acid

	H	O
Arachidic acid	1.56	n.s.
Azelaic acid	3.54	1.44
Elaidic acid	100.86	127.70
Icosenoic acid	-2.57	-4.66
Methylhexadecanoic acid	1.28	n.s.
Palmitic acid	1.38	n.s.
Stearic acid	1.55	1.51
Myristic acid	1.15	n.s.

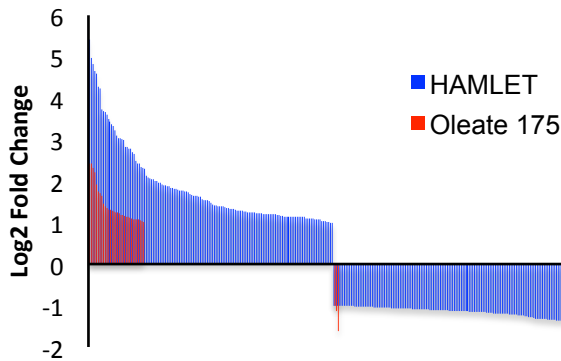
	H	O
Citric acid	-2.56	-3.44
Isocitric acid	-2.03	-3.12
a-ketoglutaric acid	-2.40	-2.17
Succinic acid	2.45	3.12
Fumaric acid	2.23	3.22
Malic acid	2.51	3.15

**Fig. 4**

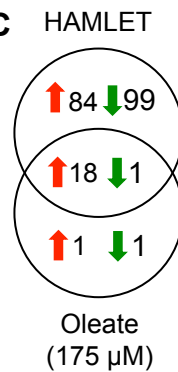
**A**



**B**



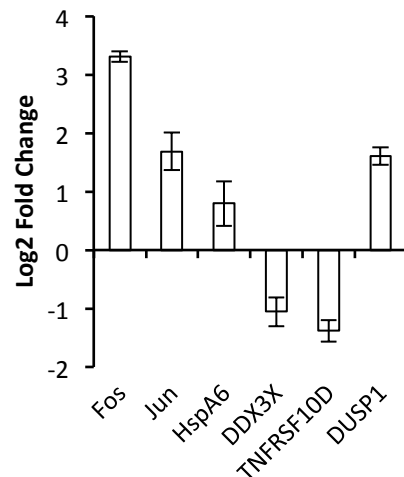
**C**



**D**

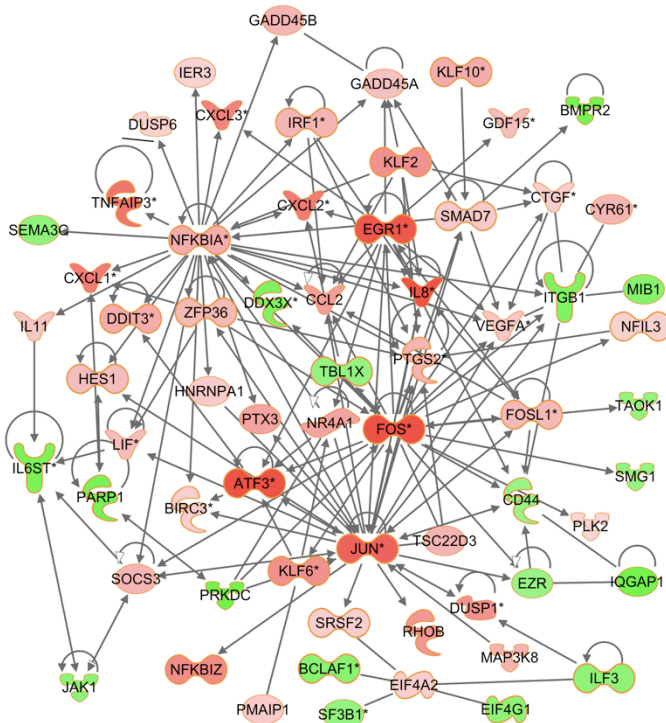
Gene set	HAMLET		Oleate 175	
	No. of genes	P Value	No. of genes	P Value
<b>Shared</b>				
MAPK signaling	13	1.17E-03	6	2.25E-04
Cytokine-cytokine receptor interaction	12	3.00E-03	4	2.10E-02
Chemokine signaling	9	1.07E-02	3	6.97E-02
<b>HAMLET/Oleate (175 μM) specific</b>				
Pathways in cancer	12	1.67E-02	-	-
NOD-like receptor pathway	7	4.67E-04	-	-
Cell cycle	6	5.25E-02	-	-
Toll-like receptor signaling	-	-	3	2.26E-02

**E**



**F**

**HAMLET**



**Oleate (175 μM)**

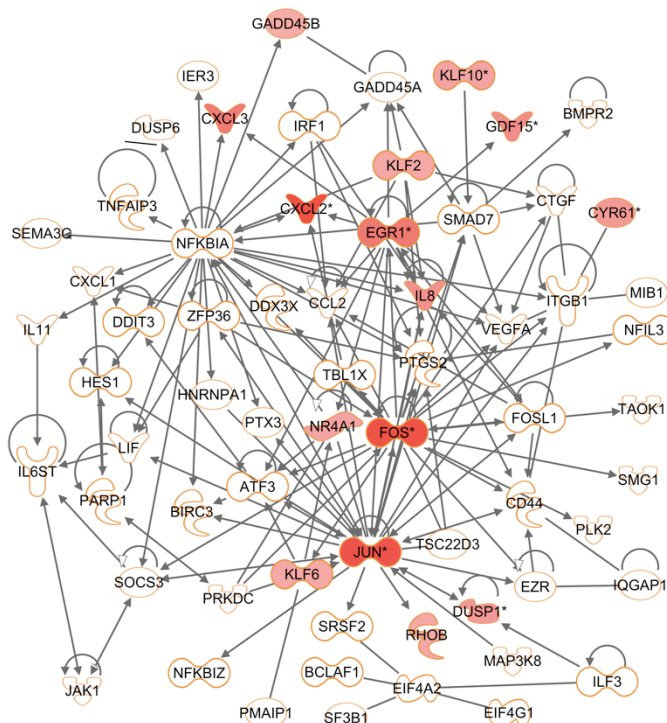


Fig. 5

