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Colonisation and molecular diversity of arbuscular mycorrhizal fungi in the aquatic plants *Littorella uniflora* and *Lobelia dortmanna* in southern Sweden

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The colonisation intensity and composition of the mycorrhizal community in the aquatic plants *Lobelia dortmanna* and *Littorella uniflora* were studied. The mycorrhizal fungi were characterised by fungal specific nested PCR and sequencing using the 5’-end of the LSU rDNA as target. For this, primers for the clade of *Acaulospora*, the clade including *Glomus mosseae* and *G. intraradices* and the clade containing *G. etunicatum* and *G. claroideum* were used. The nested PCR products were screened for different sequence types using single stranded conformation polymorphism (SSCP) and representatives for each type were sequenced. A phylogenetic analysis of the sequences showed two phylotypes of *Acaulospora*, one phylotype within the clade of *G. etunicatum/G. claroideum* and five within the *G. mosseae/G. intraradices* clade. The colonisation intensity was comparable to that seen in typical grassland vegetation. The neutral lipid fatty acid 16:1ω5 was seen to be indicative of mycorrhizal colonisation with concentrations up to 35 nmol mg⁻¹ root DW, which indicates that the fungi are active.

**INTRODUCTION**

The vegetation of oligotrophic, softwater lakes is dominated by the aquatic plants *Littorella uniflora*, *Lobelia dortmanna* and *Isoetes lacustris* (Sculthorpe 1967). They belong to the isoetids, which are characterised by a rosette growth form, a well-developed root system and an extensive aerenchyma. The occurrence of arbuscular mycorrhiza (AM) in *Lobelia* and *Littorella* was first reported in Danish oligotrophic, softwater lakes by Søndergaard & Lægaard (1977). Since then, the symbiosis has been found in aquatic and semi aquatic plants in many places in the world (Khan & Belik 1995), however only a minority of aquatic plant species have been reported to be colonised by AM fungi (Chaubal, Sharma & Mishra 1982, Nielsen, Thingstrup & Wigand 1999, Nielsen, Thingstrup & Wigand 1999, Nielsen, Thingstrup & Wigand 1999). The isoetids are adapted to nutrient uptake in the root, for example CO₂ is taken up from the interstitial water (Wiium-Andersen 1971) where it is found in 20–200-fold higher concentrations than in the overlying water (Wiium-Andersen & Andersen 1972). The permeable root surface results in oxygen leaking into the sediment (Sand-Jensen & Prahl 1982), whereby the sediment is oxygenated and the sediment redox potential (E₉ₒ) is increased (Pedersen, Sand-Jensen & Revsbech 1995). Mycorrhizal fungi are believed to be dependent on oxic conditions for their growth (Tanner & Clayton 1985, Beck-Nielsen & Madsen 2001), and the extensive aerenchyma and the oxic rhizosphere may likely explain the well-developed mycorrhiza in isoetids.

From terrestrial studies it is known that the AM symbiosis can improve uptake of phosphorus (P) and micronutrients in plants (Smith & Read 1997). The nutritional role of mycorrhiza in aquatic plants is not completely clarified, but the extensive mycorrhizal colonisation tend to correlate both with low tissue P content of the aquatic plants (Christensen & Wigand 1998) as well as with low sediment nutrient concentration (Chaubal et al. 1982). Furthermore, the low P availability in the oligotrophic and mesotrophic softwater lakes indicates a potential benefit of mycorrhiza to the plants in this ecosystem.

Presently, not much is known about the fungi forming the symbiosis with isoetids in terms of diversity.
and type of colonisation. A variety of glomalean fungi have been isolated from aquatic sediments as spores in earlier studies (e.g. Ragupathy, Mohankumar & Mahadevan 1990, Miller et al. 1999), but it is difficult to relate the presence of spores to colonisation of particular plants. PCR-based methods have a great potential for the study of diversity of AM fungi (AMF) in aquatic plant communities. The aim of the present study was to describe the mycorrhizal community and the intensity of colonisation in six softwater lakes in southern Sweden and to see if the mycorrhizal flora in aquatic plants deviated from that found in terrestrial systems. Another aim was to investigate if the mycorrhizal flora varied between the studied plant species and between the different lakes.

The large ribosomal subunit (LSU) gene was chosen as target for the PCR in this study, as this gene contains enough variation to distinguish species of AMF (Clapp, Rodriguez & Dodd 2001). The intensity of the AM colonisation was studied by use of signature fatty acids (Olsson 1999). The amount of phospholipid fatty acids (PLFA) in a given cell is well correlated to biomass (Tunlid & White 1992), whereas neutral lipid fatty acids (NLFAs) are indicators of storage carbon. We used fatty acids to describe the AMF colonisation and to compare their biomass and carbon storage levels with terrestrial ecosystems.

**MATERIALS AND METHODS**

**Collection and microscopy of plant material**

*Lobelia dortmanna* (Campanulaceae) was collected from six lakes and *Littorella uniflora* (Plantaginaceae) and *Isoetes lacustris* (Isoetaceae) from two lakes in southern Sweden (Table 1). The lakes had varying nutrient concentrations (2–111 μg l⁻¹ NO₃-N, 371–927 μg l⁻¹ tot-N, <1–2 μg l⁻¹ PO₄-P, 24–34 μg l⁻¹ tot-P; data from the Swedish Agricultural University). Three to eight specimens of each species were collected from each lake and stored in plastic bags until returned to the lab. Plant roots were pooled, cleaned and divided into three portions. Colonisation was measured by the microscopy method and by fatty acid analysis. Root samples for fatty acid analysis and molecular identification were stored at –80 °C until analysis. For microscopic analysis root samples were cleared in 10 % KOH and stained with 0.05 % aniline blue in lactoglycerol (Brundrett et al. 1996). The percentage of root length colonised and the occurrence of arbuscules and vesicles was assessed employing the method of McGonigle et al. (1990) using light microscopy at 200 times magnification. The presence of fine endophytic fungi (Thippayarugs, Bansal & Abbott 1999) was also recorded.

**Analysis of fatty acid signatures**

Phospholipid fatty acid (PLFA) composition varies between different groups of organisms, and a distinctive composition exists for the AMF. The concentration of fatty acids reflects the living biomass of fungi. The lipids from freeze-dried root samples (10–30 mg dw) were extracted according to Bligh & Dyer (1959). The lipids were separated into neutral lipids (NLFAs), glycolipids and phospholipids (PLFAs) on pre-packed silica columns (100 mg sorbent mass, Varian) by eluting with 1.5 ml chloroform, 6 ml acetone and 1.5 ml methanol, respectively. The fatty acid residues of NLFAs and PLFAs were transformed into free fatty acid methyl esters and analysed by gas chromatography using a 50 m HP5 capillary fused silica column (Hewlett Packard) with H₂ as carrier gas (Frostegård, Tunlid & Bååth 1993). The fatty acids were identified from their retention times in relation to the internal standard (FA methyl ester 19:0). These were compared to fatty acids identified using gas chromatography-mass spectrometry. We compared the NLA/PLFA ratios for the dominant fatty acids in the roots in order to determine which were of AM fungal origin. It has earlier been shown that AM fungi have large amounts of neutral FA compared with bacteria and

**Table 1.** Sampling sites and degree of mycorrhizal colonisation of *Lobelia dortmanna*, *Littorella uniflora* and *Isoetes lacustris*. TRLC, total root length colonised; AC, arbuscular colonisation; VC, vesicular colonisation (according to McGonigle et al. 1990).

<table>
<thead>
<tr>
<th>Lake</th>
<th>Lake location</th>
<th>Plant species</th>
<th>Water depth (m)</th>
<th>TRLC (%)</th>
<th>AC (%)</th>
<th>VC (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Norre sjö</td>
<td>56° 44’ N, 13° 89’ E</td>
<td><em>L. dortmanna</em></td>
<td>0.1–0.2</td>
<td>57</td>
<td>43</td>
<td>36</td>
</tr>
<tr>
<td></td>
<td></td>
<td><em>L. uniflora</em></td>
<td>0.1–0.2</td>
<td>60</td>
<td>41</td>
<td>27</td>
</tr>
<tr>
<td></td>
<td></td>
<td><em>I. lacustris</em></td>
<td>0.3</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Vittsjön</td>
<td>56° 35’ N, 13° 69’ E</td>
<td><em>L. dortmanna</em></td>
<td>0.1–0.2</td>
<td>85</td>
<td>83</td>
<td>65</td>
</tr>
<tr>
<td></td>
<td></td>
<td><em>L. uniflora</em></td>
<td>0.1–0.2</td>
<td>82</td>
<td>76</td>
<td>46</td>
</tr>
<tr>
<td>Skeingesjön</td>
<td>56° 39’ N, 13° 90’ E</td>
<td><em>L. dortmanna</em></td>
<td>0.1–0.2</td>
<td>57</td>
<td>51</td>
<td>43</td>
</tr>
<tr>
<td>Tydingen</td>
<td>56° 24’ N, 14° 03’ E</td>
<td><em>L. dortmanna</em></td>
<td>0.1–0.2</td>
<td>55</td>
<td>49</td>
<td>18</td>
</tr>
<tr>
<td>Svanshalssjön</td>
<td>56° 41’ N, 13° 94’ E</td>
<td><em>L. dortmanna</em></td>
<td>0.5–1.0</td>
<td>34</td>
<td>28</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td></td>
<td><em>I. lacustris</em></td>
<td>1.0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Osbyssjön I</td>
<td>56° 34’ N, 13° 99’ E</td>
<td><em>L. dortmanna</em></td>
<td>0.1–0.2</td>
<td>78</td>
<td>75</td>
<td>53</td>
</tr>
<tr>
<td>Osbyssjön II</td>
<td></td>
<td><em>L. dortmanna</em></td>
<td>0.1–0.2</td>
<td>66</td>
<td>57</td>
<td>50</td>
</tr>
</tbody>
</table>
Table 2. Primers used to amplify the LSU rDNA sequences from mycorrhizal roots.

<table>
<thead>
<tr>
<th>Primers</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>LSU 0061</td>
<td>5'-AGCATATCAATAAGCGGAGGA</td>
</tr>
<tr>
<td>NDL22</td>
<td>5'-TGCTCCGTTTCTCAAGACG</td>
</tr>
<tr>
<td>LSU/RK4</td>
<td>5'-GGGAGGTAATTTCCCTAAGGC</td>
</tr>
<tr>
<td>LSU/RK7m</td>
<td>5'-ATCGAAGCAGCATCTC</td>
</tr>
<tr>
<td>LSU/Aca1f</td>
<td>5'-GGGTGGTAATTTCCCTAAGGC</td>
</tr>
<tr>
<td>LSU/Aca3d</td>
<td>5'-RRWCRYCACATYYMYCARGG</td>
</tr>
<tr>
<td>LSU/Ucla1f</td>
<td>5'-GGGTGGTAATTTCCCTAAGGC</td>
</tr>
<tr>
<td>LSU/Ucla2r</td>
<td>5'-AAAGAGCCACATCTCCAG</td>
</tr>
</tbody>
</table>

R = A or G; W = A or T; Y = C or T; M = A or C.

a Primers from van Tuinen et al. (1998b).
b Primers used for primary PCR amplifications.
c Primers from Kjøller & Rosendahl (2000).
d Primers used for the second, nested PCR amplifications.
e Primers used for sequencing.

roots and therefore the NLFA/PLFA ratio for AM fungal fatty acids is normally higher than 1 and for other fatty acids lower than 1 (Olsson & Johansen 2000). It has earlier been found that the amount of NLFA 16:1ω5 in non-mycorrhizal Plantago lanceolata (a plant in the same family as L. uniflora) from pot cultures was less than 0.1 nmol mg⁻¹ dry root (van Aarle & Olsson 2003). This value was used as an indication of the presence of this NLFA in roots that was of another origin than of AM fungi. Furthermore, we compared the fatty acid composition in the AM colonised plants with the composition in the non-mycorrhizal Isoetes lacustris to verify which fatty acids could be used as indicators for AM fungal biomass.

PCR
From each sampling site and from each of the mycorrhizal plant species 16–25 randomly chosen individual root pieces of 5 mm length were transferred to 1.5 ml Eppendorf tubes and DNA was extracted by the Chelex extraction method (van Tuinen et al. 1998a, Kjøller & Rosendahl 2000). The samples were spun at 13 000 rpm for 5 min. Primary PCR was performed with the eukaryote specific primers LSU 0061 (LR1)/NDL22 (van Tuinen, Zhao & Gianinazzi-Pearson 1998b) giving an approx. 750 bp product in the variable 5'-end of the LSU rDNA (Table 2). The PCR conditions were as described by Kjøller & Rosendahl (2000).

Amplification products were used as templates in a nested PCR. For this, two new sets of primers were constructed, LSU/Ucla1f/LSU/Ucla2r specific for the Glomus claroides/G. etunicatum clade and LSU/Aca1f/LSU/Aca3r for the Acaulospora (A. longula, A. laevis, A. tuberculata, and Entrophospora colombiana) clade. A third primer set LS/URK4f/LS/URK7mr, modified after Kjøller & Rosendahl (2000), amplifying a lineage of Glomus (including G. caledonium, G. mosseae, G. geosporum) was also used (Table 2). The PCR conditions were as described by Kjøller & Rosendahl (2000). The new primers were designed from sequences available from GenBank.

Single stranded conformation polymorphism
The nested PCR products were screened for sequence variation using single stranded confirmation polymorphism (SSCP). Nested PCR products were grouped according to SSCP pattern. Representatives of each SSCP type were sequenced at the MWG Biotech facility in Germany after purification using Qiagen’s ‘Qiagen PCR purification kit’ (Quiagen, Valencia, CA). The samples were prepared as described by Kjøller & Rosendahl (2000).

In the case where SSCP analysis indicated the presence of only one sequence the nested PCR product was sequenced directly. In the case where SSCP analysis yielded complex patterns indicating that several sequences were present in the nested PCR product gel bands were cut out, squashed in 10 ml TE-buffer (Kjøller & Rosendahl 2000), and used as template in a new PCR reaction with nested primers. If the resulting pattern was similar to the original pattern it was accepted as the pattern of one sequence due to alternative conformations of this sequence. If the pattern was less complex it was assumed that the first, more complex, pattern was due to more than one sequence in the PCR product and the rest of the bands from the original SSCP gel pattern were cut out and used as PCR templates to get all the potential sequences from the original PCR product. Sequences have been deposited in the EMBL database under accession nos AJ620952–AJ620980.

Analysis of molecular data
Sequencing traces were checked for base-calling errors using the program ‘Chromas’. Sequences were compared to known sequences using the BLAST search option at NCBI (http://www.ncbi.nih.gov/index.html). Sequences that did not match with glomalean sequences were omitted from the following phylogenetic analysis.

The sequences were aligned using the computer program Clustal W, integrated in the BioEdit program package on the internet (Thompson et al. 1997) and then refined manually. The phylogenetic analysis was performed as a maximum parsimony (MP) analysis with the program PAUP (Swofford 1998), using heuristic search options with 10 000 bootstrap replications. The settings for the analysis were: branch swapping by tree-bisection-reconnection (TBR), branches able to collapse to yield polytomies and with parsimony uninformative characters excluded (Clapp et al. 2001). The phylogenetic trees were rooted with Paraglomus occultum as an out-group (Morton & Redeker 2002). Using the information in a strict consensus tree ‘phylotypes’ (Vandenkoornhuyse et al. 2002) were assigned to clades of closely related
sequences that had high statistical support by bootstrap analysis.

RESULTS

AM fungal colonisation and signature fatty acids

The AMF colonisation of *Lobelia* and *Littorella* were from 34–85% of the total root length, the range of arbucular colonisation was 28–83%, and vesicular colonisation 6–65% (Table 1). The lowest colonisation was found in plants from Lake Svanshalssjön where plants were sampled at the largest depth. *Isoetes* was not colonised by AM fungi (Table 1). Intraradical spores were recorded in *Lobelia* collected in Lake Osbyssjön (site II). Colonisation by fine endophytic fungi was recorded in *Lobelia* in two of the lakes (Lake Norre sjö and Lake Skeingesjön). *Ranunculus reptans* and *Hydrocotyle vulgaris* were found in semiaquatic environments in some of the lakes and were also found to be AM colonised. *Hydrocotyle* showed Paris-type of colonisation, while the other sampled species had colonisation of Arum-type (Smith & Smith 1997).

The most common 18C and 16C fatty acids identified in roots of the collected aquatic plants were 16:0, 16:1o5, 16:1o7, 18:0, 18:1o7, and 18:2o6,9. There was no difference in total PLFA content between the different plants, while the two AM species had significantly higher NLFA contents than the non-mycorrhizal *Isoetes* (Table 3). AM fungi have a high NLFA to PLFA ratio and this ratio for different fatty acids had a ratio lower than one. All fatty acids had a ratio lower than one in the non-mycorrhizal *Isoetes*. The content of the signature NLFAs was in general higher in *Littorella* than in *Lobelia*. The NLFA concentrations for the signature fatty acids in *Lobelia* and *Littorella* were many times higher than in the non-mycorrhizal *Isoetes*, and also several orders of magnitude higher than what have been reported for non-mycorrhizal control plants in pot experiments (van Aarle & Olsson 2003). The content of NLFA 16:1o5 in *Lobelia* was highest in Lake Osbyssjön (35 nmol mg⁻¹), whereas the lowest content was found in Lake Tydingen (0.15 nmol mg⁻¹).

<table>
<thead>
<tr>
<th>Fatty acid content nmol mg⁻¹ root</th>
<th><em>L. uniflora</em> (n = 2)</th>
<th><em>L. dortmanna</em> (n = 8)</th>
<th><em>I. lacustris</em> (n = 2)</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Total PLFA</strong></td>
<td>15.9 ± 1.6</td>
<td>13.6 ± 1.2</td>
<td>14.9 ± 3.0</td>
<td>*</td>
</tr>
<tr>
<td><strong>Total NLFA</strong></td>
<td>37.9 ± 9.7</td>
<td>19.5 ± 9.1</td>
<td>2.3 ± 0.4</td>
<td>*</td>
</tr>
<tr>
<td><strong>NLFA/PLFA</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>16:0</td>
<td>2.24 ± 0.30</td>
<td>1.31 ± 0.44</td>
<td>0.12 ± 0.01</td>
<td>*</td>
</tr>
<tr>
<td>16:1o5</td>
<td>79.6 ± 13.3</td>
<td>25.8 ± 9.7</td>
<td>0.40 ± 0.01</td>
<td>**</td>
</tr>
<tr>
<td>16:1o7</td>
<td>2.30 ± 0.54</td>
<td>2.38 ± 0.48</td>
<td>0.15 ± 0.01</td>
<td>***</td>
</tr>
<tr>
<td>18:0</td>
<td>0.69 ± 0.02</td>
<td>0.55 ± 0.10</td>
<td>0.39 ± 0.11</td>
<td></td>
</tr>
<tr>
<td>18:1o7</td>
<td>0.35 ± 0.05</td>
<td>0.88 ± 0.21</td>
<td>0.59</td>
<td></td>
</tr>
<tr>
<td>18:2o6,9</td>
<td>0.31 ± 0.05</td>
<td>0.65 ± 0.16</td>
<td>0.17 ± 0.03</td>
<td></td>
</tr>
<tr>
<td><strong>NLFA</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>16:0</td>
<td>12.3 ± 3.8</td>
<td>5.8 ± 2.5</td>
<td>0.78 ± 0.16</td>
<td>*</td>
</tr>
<tr>
<td>16:1o5</td>
<td>21.9 ± 5.1</td>
<td>7.5 ± 4.8</td>
<td>0.04 ± 0.03</td>
<td>**</td>
</tr>
<tr>
<td>16:1o7</td>
<td>0.28 ± 0.04</td>
<td>0.57 ± 0.19</td>
<td>0.05 ± 0.01</td>
<td>*</td>
</tr>
</tbody>
</table>

Fungal diversity

DNA was extracted from 202 root pieces of which 181 (90%) gave a primary PCR product. Of these, 117 (64%) gave a nested PCR product with one or more primer sets (Table 4). In all, 50 nested PCR products from *Littorella* and 104 from *Lobelia* were subject to SSCP analysis. PCR products with the LSU RK4f/RK7nr primers yielded 9 different SSCP patterns indicating potentially 9 different sequences. Three different patterns were identified with the primers LSU claf/clar2r and 4 different patterns with the LSU Aca1f/Aca3r primers, in all 16 potential patterns were distinguished. Representatives for each pattern were sequenced; in all 29 nested PCR products were sequenced. 16 sequences were most closely related to the clade containing *Glomus mosseae*, seven to the clade of *G. etunicatum/G. claroideum*, and six to the *Acaulospora* clade. The 29 root derived glomalean sequences were used in a maximum parsimony analysis comparing them to earlier obtained spore and root-derived sequences in the Department of Mycology (University of Copenhagen) and from the NCBI-GenBank database (Fig. 1). The phylogenetic analysis grouped the sequences into eight different phylogenotypes, meaning that the root systems analysed potentially contained eight distinct clades of AM fungi.
 taxa within the families studied. Eleven sequences fell in two closely related clusters between G. intraradices and the clade containing G. caledonium and G. mosseae. These 2 clades were named phylotype 1 and 2 (Fig. 1). Five of the sequences clustered with root derived Glomus sp. sequences found by S. R. in Hieracium pilosella (data not shown). These were named phylotype 3, 4, and 5. Seven sequences clustered to form a clade with some sequences from G. claroideum isolates and were distinguishable from the clade containing isolate BEG88. This clade was termed phylotype 6. Six sequences clustered within the Acaulospora clade, five in one distinct clade close to A. longula, that we named phylotype 7; the sixth sequence was named phylotype 8.

Both Lobelia and Littorella harboured sequences targeted with all three primer-sets. Phylotypes 1, 6, and 7 were found in high numbers in both plant species, phylotype 5 was only found in Littorella, phylotypes 2, 3, and 8 were only found in Lobelia (Fig. 2). The number of phylotypes encountered for one plant species in one lake varied from 2–6. Looking at the distribution of phylotypes in different lakes (Fig. 2) phylotypes 1, 6, and 7 were in general present at high frequencies. Three lakes were atypical in that they lacked one of these common phylotypes. In Lake Svanhallssjön no Acaulospora sequences were seen (out of 14 sequences). In Lake Vittsjön no sequences from the clade of G. etunicatum/G. claroideum were seen (out of 13 sequences). In Lake Skeingesjön the most dominant phylotype in the clade containing G. mosseae and G. intraradices, phylotype 1, was not found. Only one sequence from this clade was encountered in this lake (phylotype 4, accounting for 7% of the found sequences). For the one lake where we sampled at two places (100 m apart), Lake Osbysjön, a comparable phylotype composition was seen at both places.

**DISCUSSION**

The dense vegetation of isoetids often found in temperate, oligotrophic, softwater lakes plays an important role in keeping the lakes in an oligotrophic state (e.g. Andersen & Olsen 1994, Olsen & Andersen 1994). The isoetid vegetation has been dramatically reduced in lakes in Northern Europe and Northern America during the last century (Arts 2002). A better understanding of the ecology of isoetids is therefore needed. The heavy mycorrhizal colonisation found in some isoetids indicates that this symbiosis may be important for many members of this plant group. There are indications of the importance of mycorrhiza for the phosphorus supply of isoetids (Christensen & Wigand 1998). However, there is a big gap in our knowledge regarding the influence of diversity in AMF, studied as molecular variation, on the function of these in the aquatic plants. This subject is important for understanding the ecology of isoetids.

The phylotype composition of the six lakes studied was quite similar although the lakes were situated up to 25 km apart and had a rather broad span of nutrient concentrations. For some of the lakes we had a relatively low success rate of the nested PCR (Table 4), which was independent of the degree of colonisation. It is likely that not all fungi observed by microscopy were viable or that the observed fungi were for some part of other families than the ones targeted with our primers, which would explain this observation. The number of phylotypes found varied among lakes from two (Lobelia in Lake Vittsjön) to six (Lobelia at Lake Osbysjön II) and was not dependent on the number of sequences found. In Lake Tydingen we found five different phylotypes with only seven sequences, and in Lake Vittsjön only two phylotypes were found although thirteen sequences were obtained.

A comparison of the phylogeny presented by Schwarzott, Walker & Schüssler (2001) based on the SSU with the one presented here (Fig. 1) shows that both clearly differentiate between the groups containing Glomus mosseae, with G. intraradices, and G. claroideum/G. etunicatum. This shows that the two datasets are comparable. The 5' LSU region is more variable than the SSU region (van Tuinen et al. 1998a), and using the LSU region may identify additional phylotypes than the SSU region would have identified from the same material. Bearing this in mind we compared the diversity of RFLP types in woodland and farmland

<table>
<thead>
<tr>
<th>Plant species, Lake</th>
<th>RP</th>
<th>1° PCR</th>
<th>Success rate (%)</th>
<th>LSU/URK4f/7m</th>
<th>LSU/claf/2r</th>
<th>LSU/Acaf/3r</th>
<th>AM sequences</th>
</tr>
</thead>
<tbody>
<tr>
<td>L. dortmannana, Norre sjö</td>
<td>16</td>
<td>10</td>
<td>40</td>
<td>3</td>
<td>2</td>
<td>3</td>
<td>7</td>
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<tr>
<td>L. dortmannana, Vittsjön</td>
<td>20</td>
<td>16</td>
<td>50</td>
<td>5</td>
<td>0</td>
<td>8</td>
<td>13</td>
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<tr>
<td>L. dortmannana, Skeingesjön</td>
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<td>17</td>
<td>59</td>
<td>7</td>
<td>4</td>
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<tr>
<td>L. dortmannana, Tydingen</td>
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<td>18</td>
<td>33</td>
<td>6</td>
<td>1</td>
<td>4</td>
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<tr>
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<td>42</td>
<td>31</td>
<td>23</td>
<td>2</td>
<td>3</td>
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<tr>
<td>L. dortmannana, Osbysjön I</td>
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<td>21</td>
<td>67</td>
<td>13</td>
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<tr>
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<td>7</td>
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<tr>
<td>L. uniflora, Norre sjö</td>
<td>20</td>
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<td>9</td>
<td>8</td>
<td>6</td>
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<tr>
<td>L. uniflora, Vittsjön</td>
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<td>60</td>
<td>11</td>
<td>8</td>
<td>8</td>
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</tbody>
</table>

Table 4. Number of root pieces (RP) analysed, number of RP yielding primary PCR product, success rate of nested PCR, number of sequences with LSU/URK4f/7m primers, LSU/claf/2r primers, LSU/Acaf/3r primers, total number of sequences examined, number of AM LSU gene sequences found, listed for each lake studied.
Acaulospora G. claroideum clade

LobRK183
LobRK147
LitRK210
LobRK204

Hp, Glomus seq.1 (AJ634603)

LitRK225
Pea root, Glomus seq.Bm (AF236737)

G.caledonium BEG20
Pea root, Glomus seq.Cl (AF236744)

G.geosporum BEG90
Pea root, Glomus seq.Dg (AF236751)

G.constrictum BEG130
Pea root, Glomus seq.Ac (AF236718)

G.mosseae BEG83

G.mosseae BEG25

G.mosseae clade

G.etunicatum BEG137

G.claroideum BEG88

G.etunicatum BEG92

G.claroideum BEG14

G.claroideum BEG89

G.claroideum BEG31

G.claroideum BEG3

Lobcla205
Litcla229
LobRK195
LobRK118
Litcla214
LobRK198

A.spinosa BEG10

A.longula (AF389006)

L. longula (AF389007)

5 changes

Fig. 1. A maximum parsimony phylogenetic tree showing AM fungal sequences isolated from roots of Lobelia dortmanna (Lob) and Littorella uniflora (Lit) using the LSURK4f/RK7mr (Glomus mosseae clade), LSUcla1f/cla2r (G. claroideum/etunicatum clade), and LSUAca1f/Aca3r (Acaulospora) primers. Bootstrap values are from the majority rule consensus tree (10,000 replicates). Values are shown where they exceed 80%. Branches present in a strict consensus tree are drawn as fat. Sequences from this study are in bold and the names are composed of abbreviations of plant species name (Lob = Lobelia, Lit = Littorella), primer (Glo = LSURK4f/RK7mr, cla = LSUcla1f/cla2r, Aca = LSUAca1f/Aca3r) and root piece number extracted. Other names are A = Acaulospora; G = Glomus, followed by strain name, BEG code or accession no. from NCBI. Pea root derived sequences are from Kjøller & Rosendahl (2001). The sequences named Hp are from colonised roots of Hieracium pilosella from an ongoing field study in Denmark.
(Helgason et al., 1998) in Yorkshire (determined from the SSU gene) to the diversity of phylotypes in the present study (determined from the LSU gene). It seems that the lakes contained a higher diversity than what was found for farmland where the sequence type called Glo1 (containing sequences of G. mosseae) was dominant, with only one Scutellospora and one Acaulospora sequence found out of 100 sequences. The diversity of phylotypes we saw in the lakes was more similar to the woodland, which had a higher diversity than the farmland, with a higher number of sequences in several different clades. In comparison to the studies by Vandenkoornhuyse et al. (2002) in permanent grassland in Scotland, both the lake, woodland and farmland sites mentioned above seem to have a rather low diversity even when disregarding

Fig. 2. The frequency of AM fungal phylotypes in Littorella uniflora and Lobelia dortmanna both as a total of all lakes sampled and for each of the seven studied locations in southern Sweden. ‘n’ denotes the number of sequences used for each graph.
Scutellospora sequences, a clade not studied in our work.

Within the clades investigated we saw that the fungi forming symbiosis with Lobelia and Littorella in lakes in southern Sweden are not so different from already known sequences of glomalean fungi from terrestrial plants (Fig. 1). It is accepted that aquatic angiosperms have evolved from terrestrial angiosperms (Cook 1990) and it is likely that the fungi and plants recolonised the aquatic habitat together. From the present data we neither prove nor disprove that some of the identified phylotypes are specific to the aquatic environment. For this, reciprocal tests of AMF cultures originating from terrestrial and from the aquatic environment with plants from both environments are needed.

The variation of AMF sequences found in Lobelia and Littorella are similar (Fig. 2). The phylotypes only encountered in Lobelia (nos 2–3 in the G. mosseeae clade, and 8 in Acaulospora) were found in lakes where Littorella was not sampled (Fig. 2). We can therefore not conclude that these are specific for Lobelia. Phylotype 5 was only found in Littorella in Lake Vittsjoen.

None of the phylotypes found in the G. mosseeae/ G. intraradices clade clustered closely with known Glomus species. The sequences constituting phylotype 1 and 2 were positioned between G. intraradices and the clade of G. caledonium, G. mosseeae, G. constrictum, G. coronatum, and G. geosporum (Fig. 1). Phylotypes 3, 4, and 5 clustered with root-derived sequences from field samples of Hieracium pilosella L. (abbreviated Hp in Fig. 1) from eastern Denmark (S.R. unpubl.). The lack of named sequences in some clades could be due to incomplete sequencing of known Glomus species; these clades could represent non-described Glomus species or even non-sporulating and therefore also non-described Glomus species. Our sequences in phylotype 6 clustered closely with G. claroideum strain BEG3 and the sequences could not be differentiated from G. claroideum strain BEG14, BEG89, and V284 in the strict consensus tree and are thus identified as G. claroideum sequences. Phylotype 7, in the Acaulospora clade, is very close to Acaulospora longula (accession nos AF389006, and AF389007 at NCBI, Fig. 1). Phylotype 8 (LobAca183, Fig. 1) groups with an A. spinosa sequence. Large ribosomal subunit gene sequences are not well represented in the NCBI database as yet and the designation of species to the rest of the phylotypes must await the sequencing of more named isolates for comparison.

The high AM colonisation in Lobelia and Littorella is similar to the intensity found in typical P limited terrestrial grasslands. For example Plantago lanceolata, which also belongs to the family Plantaginaceae, had 60% of the root length colonised in limestone habitats in southern Sweden but only 20% in acid rock habitats where the availability of P is much higher than in limestone habitats (van Aarle, Soderstrom & Olsson 2003). Isoetes lacustris, a seedless vascular plant belonging to the division Microphylophyta, did not show any sign of AM colonisation but has earlier been reported both as non-mycorrhizal (Sondergaard & Lægaard 1977) and mycorrhizal (Farmer 1988, Sharma 1998, Wigand et al. 1998) in different studies. None of these studies, however, have reported the occurrence of arbuscules. Because no mycorrhizal structures were observed in Isoetes roots by microscopic examination in this study, we used the Isoetes plants as non-mycorrhizal reference in the fatty acid analysis together with data from earlier studies on the concentration in non-mycorrhizal roots.

AM fungi have a high NLFA to PLFA ratio and this ratio for different fatty acids may thus indicate if they are AM fungal signatures. That the fatty acids 16:0, 16:1o5, and 16:1o7 had NLFA/PLFA ratios higher than one, while the other fatty acids had a ratio lower than one, together with the fact that all fatty acids had a ratio lower than one in the non-mycorrhizal Isoetes, shows that these three fatty acids do indicate mycorrhizal colonisation. The results in this study, together with earlier studies (Olsson 1999) shows that NLFA 16:1o5 is indicative of mycorrhizal colonisation and therefore we used it as indicator of AM fungal storage lipids. It showed that the amount of AM fungal storage lipids was in some cases higher than what has earlier been obtained using Glomus in greenhouse studies. Cucumber roots colonised with Glomus invermaium had 15 nmol NLFA 16:1o5 g⁻¹ (Olsson et al. 1995), while up to 52 nmol NLFA 16:1o5 g⁻¹ was found in an experiment with Plantago lanceolata (van Aarle & Olsson 2003). In field grown linseed with around 40% of the root length colonised, the amount was only 1 nmol 16:1o5 g⁻¹ (Olsson et al. 1999). This shows that the AM fungal colonisation in several of the lakes was vital in the sense that the carbon nutrition of the AM fungi was at least as successful as in many terrestrial systems. Earlier work by Olsson and others has shown that hyphal biomass correlates with 16:1o5 (Olsson 1999). There was, however, no correlation observed between colonisation intensity and the NLFA 16:1o5 in this study (results not shown). It should be noted that the staining for microscopic examination stains both living and dead fungi, whereas the fatty acids represents living biomass. This is illustrated by the fact that the biomass indicators (NLFA) correlated with the number of sequences found for a root sample (R²=0.6).

This study represents the first step towards describing the diversity of AMF found in aquatic plants. The data show that both Lobelia and Littorella have similar dominating AM fungi throughout the range of the six lakes studied. We found that the AM fungal community in the aquatic environment was not composed of a low number of specific fungi. Instead, the diversity was just as high as in terrestrial communities so far investigated.
Molecular diversity of AM fungi in aquatic plants

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REFERENCES


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