Androgens Inhibit the Stimulatory Action of 17β-Estradiol on Normal Human Breast Tissue in Explant Cultures.

Eigeliene, Natalija; Elo, Teresa; Linhala, Mari; Hurme, Saija; Erkkola, Risto; Härkönen, Pirkko

Published in:
Journal of Clinical Endocrinology and Metabolism

DOI:
10.1210/jc.2011-3228

2012

Link to publication

Citation for published version (APA):

Total number of authors:
6
Androgens Inhibit the Stimulatory Action of 17β-Estradiol on Normal Human Breast Tissue in Explant Cultures

Natalija Eigēlienē, Teresa Elo, Mari Linhala, Saija Hurme, Risto Erkkola, and Pirkko Härkönen

Department of Cell Biology and Anatomy (N.E., T.E., M.L., P.H.), Institute of Biomedicine, University of Turku, and Department of Obstetrics and Gynecology (N.E., R.E.), Turku University Central Hospital, 20520 Turku, Finland; Department of Biostatistics (S.H.), University of Turku, 20014 Turku, Finland; and Department of Laboratory Medicine (P.H.), Tumor Biology Unit, Lund University, SE-20502 Malmö, Sweden

Background: The data concerning the effects and safety of androgen in human breast tissue are conflicting.

Objective: Our aim was to analyze the effects of androgens on normal human breast tissue (HBT).

Approach: We cultured explants of HBT (obtained from reduction mammoplasty operations of postmenopausal women) with or without testosterone (T) and 5α-dihydrotestosterone (DHT) or in combination with 17β-estradiol (E2) for 7 and 14 d to study the effects of androgens on proliferation, apoptosis, target gene expression, and steroid receptors. The androgen receptor (AR) and estrogen receptor (ER) dependences of the effects were studied with the antihormones bicalutamide and fulvestrant, respectively.

Results: The hormone responsiveness of cultured breast tissue was assessed by assaying apolipoprotein-D and prostate-specific antigen expression increased by androgens and amphiregulin and trefoil factor-1 expression induced by E2 treatment. T and DHT reduced proliferation and increased apoptosis in breast epithelium, the effects of which were reversed by bicalutamide. In combination with E2, they suppressed E2-stimulated proliferation and cell survival. DHT also inhibited basal \((P < 0.05)\) and E2-induced expression of cyclin-D1 mRNA \((P < 0.05)\). Immunohistochemistry showed that T \((P < 0.05)\) and DHT \((P < 0.05)\) increased the relative number of AR-positive cells, whereas ERα-positive \((P < 0.001)\) cell numbers were strongly decreased. The percentage of ERβ-positive cells remained unchanged. E2 treatment increased ERα-positive \((P < 0.01)\) cells, whereas AR- \((P < 0.05)\) and ERβ-expressing \((P < 0.001)\) cells diminished. These effects were repressed in combination cultures of E2 with T and DHT.

Conclusion: T and DHT inhibited proliferation and increased apoptosis in the epithelium of cultured normal HBT and opposed E2-stimulated proliferation and cell survival in an AR-dependent manner. These effects were associated with changes in the proportions of ERα- and AR-positive epithelial cells. (J Clin Endocrinol Metab 97: 0000–0000, 2012)

Androgens perform physiologically important functions in females, but their effects on mammary tissue and their role in the development of breast cancer remain unclear (1, 2). There are conflicting reports on the inhibitory effects of androgens on the proliferation of breast epithelial cells, which may play a protective role in the pathogenesis of breast cancer (3). Epidemiological studies demonstrate that high levels of endogenous androgens in...
postmenopausal women are associated with an increased risk of breast cancer, independent of circulating estrogen levels (4). Experimental data from animal studies and cell cultures with human breast cancer cell lines have indicated both proliferative (5) and antiproliferative (6) effects on cell growth. Recently the presence of an androgen receptor (AR) in tumor tissue was found to be correlated with favorable outcomes in patients (7), which suggests a protective role for AR signaling in breast tissue.

Most in vitro studies using breast cancer cell lines show that androgens inhibit the proliferative effects of estrogen (8, 9). However, the results diverge, depending on the specific cell line, the status and type of steroid receptor, and the dose of androgen (10). Rodent models have limitations because of the marked biological differences between the species, which limits the validity of the data. Studies on nonhuman primates with similarities to humans in terms of the mammary gland, reproductive physiology, and peripheral steroid hormone metabolism showed that the administration of testosterone markedly reduced estrogen-induced proliferation and estrogen receptor (ER)-α gene expression (6, 11). Female-to-male transsexuals treated with androgens exhibited involutionary changes similar to those observed in the breasts of menopausal women, including marked reduction of glandular tissue, prominence of fibrous connective tissue, and the presence of only small amounts of adipose tissue (12). However, the doses of androgens used in those treatments were notably high. Thus, further clarification of the effects of androgens on human breast tissue (HBT) is required.

In this study, we aimed to clarify the action of exogenous androgens on normal HBT. The effects of testosterone (T) and 5α-dihydrotestosterone (DHT) and in combination with 17β-estradiol (E2) on HBT were studied using a modified explant culture system that allows the maintenance of epithelial-stromal interactions and the monitoring of the responses of HBT to steroid hormones (13).

Subjects and Methods

HBT sampling

HBT samples were collected from postmenopausal women (n = 19) aged 46–64 yr undergoing reduction mammoplasties at Turku University Central Hospital (Turku, Finland). None of the women had used hormonal therapy within the previous 6 months. Informed written consent was obtained from all of the patients before surgery. The study protocol was approved by the Joint Ethics Committee of Turku University and Turku University Central Hospital.

Organ culture

HBT were processed for explant culture as described previously (13) (Supplemental Methods, published on The Endocrine Society’s Journals Online web site at http://ecm.endojournals.org). 17β-Hydroxy-3-oxo-4-androstenone (T; Fluka Chemie AG, Buchs, Switzerland), 5α-androstan-17β-ol-3-one (DHT; Fluka-Chemie), and E2 (Sigma Aldrich GmbH, Steinheim, Germany), or a combination thereof was added to the basal medium. T and DHT were used at a concentration of 1 nM. E2 was used at a final concentration of 10 nM. The steroids were dissolved in dimethyl sulfoxide (Sigma). The HBT samples were collected on d 7 and 14 of the culture for further analyses [histology, whole mounts, immunohistochemistry, Western blots, and quantitative RT-PCR (qRT-PCR)]. To study the AR- and ER-mediated effects of androgens and E2, the antiandrogen bicalutamide (Bclt) and the antiestrogen fulvestrant (Fulv) were used (both kindly provided by Hormos Medical, Turku, Finland) at concentrations of 1 μM.

Whole mounts

HBT samples were spread onto a glass slide, fixed in a 1:3 mixture of glacial acetic acid/absolute ethanol, hydrated, stained overnight in 0.2% carmine and 0.5% AlK(SO4)2, dehydrated in graded solutions of ethanol, and cleared in xylene.

Morphology and immunohistochemistry

For histology and immunohistochemistry, the samples were fixed in 4% paraformaldehyde, and paraffin sections (4 μm) were mounted on SuperFrost +Plus slides (Menzel Gmbh & Co. KG, Braunschweig, Germany). The sections were deparaffinized in xylene and rehydrated through graded ethanol solutions. Endogenous peroxidase activity was blocked using 0.3% H2O2 for 30 min. The sections were then incubated with primary antibodies (Supplemental Table 1) overnight at 4°C followed by incubation with biotin-labeled secondary antibody diluted 1:200 for 1 h at room temperature. Immunoreaction was detected using the Vectastain Elite ABC kit (Vector Laboratories, Burlingame, CA) and peroxidase substrate 3,3-diaminobenzidine (Vector Laboratories) following the manufacturer’s instructions. The samples were subsequently counterstained with Mayer’s hematoxylin and mounted in Mountex (Histolab Products AB, Göteborg, Sweden).

Image analysis and histomorphometry

The sections were interpreted under light microscopy in high-power fields. An Olympus BX-60 (Olympus Deutschland GmbH, Hamburg, Germany) microscope with CellD imaging system connected to a Leica DC300F camera (Leica Microsystems GmbH, Heidelberg, Germany) was used for the image analysis. For each sample, more than 1000 common epithelial cells were scored for the various markers, and the data were expressed as the percentage of cells that were positive.

A histomorphometric evaluation was performed to estimate the changes of epithelial morphology (acinar and ductal wall thickness) during different hormonal treatments using OsteoMeasure system (OsteoMetrics, Atlanta, GA) as described previously (13) (Supplemental Methods).

Western blot analyses

Cultured frozen HBT was homogenized in lysis buffer (150 mM NaCl, 50 mM Tris-HCl, pH 8.0). The protein concentration was determined using the Bio-Rad protein assay kit (Bio-Rad Labora-
for 5 min in Laemmli sample buffer with 2β-mercaptoethanol and loaded onto a 12% SDS-PAGE gel. After gel electrophoresis, the proteins were transferred to a polyvinylidene fluoride membrane (Bio-Rad Laboratories). The primary antibodies used for the Western blots are presented in Supplementary Table 1. The secondary goat antimonium IgG-horseradish peroxidase (Bio-Rad Laboratories) was used at a dilution of 1:10,000. Detection was performed using ECL-Plus (Amerham, Piscataway, NJ). The band intensities were quantified using Analysis MCID version 5.0 software (Imaging Research, Inc., Ontario, Canada), and the results are expressed as relative density of target to tubulin (loading control) ratios.

**Quantitative RT-PCR**

Total RNA from the frozen HBT was isolated using the TRIzol reagent (Invitrogen, Carlsbad, CA) according to the manufacturer’s instructions. One microgram of total RNA was reverse transcribed with AMW reverse transcriptase (Finnzymes, Espoo, Finland). The cDNA was diluted 1:10 in nuclelease-free water, and 8 μl was analyzed in duplicate by RT-PCR in a CFX384 RT-PCR detection system (Bio-Rad Laboratories) using the DyNAmo SYBR Green qPCR kit (Finnzymes) with 0.4 μmol/liter of each primer (Oligomer, Helsinki, Finland) (Supplementary Table 2) in a reaction mixture with a total volume of 20 μl. The negative controls contained water in place of first-strand cDNA. The relative gene expression levels were normalized to a calibrator that was chosen to be the control group. The final results were expressed as n-fold differences in gene expression relative to keratin-18 and the control. The results were calculated using the cycle threshold (Ct) method as follows: n-fold = \( 2^{-\Delta \Delta Ct} \), where the ΔCt values of the sample and the controls were determined by subtracting the average Ct value of the housekeeping gene from the average Ct value of the different genes analyzed.

**Statistical analysis**

Statistical analysis was performed using general linear mixed models. The effect of the treatment type and time were also tested. The Tukey-Kramer method was used to adjust the P values of the pairwise comparisons, and the residuals were checked for justification of the analysis. The data are expressed as mean ± sd. P < 0.05 were considered to be statistically significant. Statistical analyses were carried out using the IBM SPSS Statistics 19 system (IBM Corporation, White Plains, NY).

**Results**

**Effects of androgens on HBT morphology**

For the control HBT cultured for 7 or 14 d, the explants maintained their structural integrity and glandular morphology. The epithelial cells were cuboidal and were arranged in two regular layers. Pilot testing was done to determine the optimal concentration for studying the biological effects of androgens on HBT. The concentration of 1 nM T and 1 nM DHT were used in further experiments alone or in combination with E2.

The 10-nm concentration of E2 was used based on previous studies, which determined effective concentrations of these steroid hormones in vitro (11). As previously shown (13), HBT was strongly stimulated with E2. The acinar epithelium became stratified, and a portion of luminae was obliterated. The epithelial cells varied in size and shape, and they appeared to lose their polarized orientation in relation to the basement membrane, especially in the explants grown with E2 for 14 d (Fig. 1A, upper panel). A combination of E2/T or E2/DHT prevented the E2-induced changes. Similar to the controls, the epithelium remained cuboidal, regular and organized, and the epithelial cells retained their polarity and height. The histomorphometry showed that the thickness of acinar and ductal wall was increased by treatment with E2 and decreased by T and DHT significantly (Fig. 1A, lower panel). Whole mounts confirmed the involutive effects of DHT and the stimulatory effects of E2 on the lobule structure of explants cultured for 14 d (Fig. 1B).

**Hormone responsiveness of androgen and E2 target gene expression in cultured HBT**

Hormone responsiveness was examined by studying the expression of androgen-regulated prostate-specific antigen (PSA) and apolipoprotein D (Apo-D) (14) vs. E2-regulated trefoil factor-1 (TFF1) (14) and amphiregulin (AREG) (15, 16) in HBT grown with or without T or DHT vs. E2, respectively. Immunohistochemistry showed that PSA stained faintly in the myoepithelial cells of the control samples, whereas it was strongly expressed in the epithelial cells of HBT cultured with androgens (Fig. 2A, left panel). In the presence of Bclt, T/Bclt, or DHT/Bclt, the PSA immunoreaction was low. The quantification of PSA-positive cells showed that androgens increased the percentage of PSA-positive cells compared with the controls (Fig. 2A, right panel). Bclt decreased the PSA-positive cell proportion to approximately 25% and significantly attenuated the effects of T and DHT. Treatment with E2 and Fulv had a negative effect on the percentage of PSA-positive cells. Fulv decreased the PSA-positive cell count to approximately 30%. The addition of E2 to Fulv significantly increased the percentage of PSA-positive cells, but their level remained lower than in the control groups. Western blot analyses showed that T and DHT increased the expression of Apo-D and their effects were blocked by Bclt (Fig. 2B).

TFF1 expression was strong in the epithelial cells of explants cultured with E2 (Fig. 2C, left panel). In the Fulv-treated explants, TFF1 was observed only in clumps of detached epithelial cells located in the luminae and not in the epithelium. The addition of Fulv with E2 diminished TFF1 staining in the epithelial cells, but there were TFF1 cell clumps in some luminae. Treatment with E2 increased the level of TFF1 mRNA up to 6-fold compared with the
controls ($P < 0.001$) (Fig. 2C, right panel). Fulv decreased the basal and E$_2$-induced TFF1 mRNA. Both androgens inhibited TFF1 mRNA expression, and Bclt alone increased it. Interestingly, the addition of T to Bclt further increased TFF1 mRNA compared with the control, whereas the addition of DHT to Bclt reversed TFF1 mRNA to the control level (Fig. 2C). The Western blotting experiments showed that E$_2$ increased the level of AREG in HBT (Fig. 2B), an effect opposed by Fulv.

Androgens decrease epithelial cell proliferation in HBT

The proliferation was determined by immunohistochemistry with phosphohistone-3 (PHH3) (Fig. 3A) and Ki67 antigen (Fig. 3B). The relative numbers of PHH3-positive cells were lower compared with those of the Ki67-positive cells, reflecting the expression pattern of the markers during the cell cycle (17). The number of PHH3 cells was lower in T- ($P < 0.05$) and DHT-treated ($P < 0.001$) HBT compared with the controls (Fig. 3A). Interestingly, the numbers of PHH3-positive cells increased in Bclt-treated HBT ($P < 0.01$) compared with the controls. The addition of Bclt to T or DHT opposed these androgen effects (Fig. 3A). The highest percentage of PHH3-positive cells was observed in the E$_2$ group when compared with the controls and other treatments (Fig. 3A, right panel). Similar results were obtained with Ki67 (Fig. 3B). The effects of E$_2$/T and E$_2$/DHT on proliferation were assessed only with Ki67 (Fig. 3C). As previously shown (13), the proliferation increased in the E$_2$-treated HBT ($P < 0.01$) and decreased in a time-dependent manner in HBT treated with T and DHT for 7 and 14 d (Fig. 3C, right panel). The combination of T and DHT with E$_2$ opposed the E$_2$-alone effects as compared with the HBT cultured with E$_2$ alone at both 7 and 14 d.

Expression of cyclin-D1 in HBT cultured with androgens and E$_2$

To study the mechanisms of androgen inhibition of HBT proliferation in the presence and absence of E$_2$, the expression of cyclin-D1 was studied by immunohistochemistry and qRT-PCR (Fig. 4). Approximately 10% of the epithelial cells were immunopositive for cyclin-D1 in the controls, whereas a larger proportion was observed in the E$_2$-treated explants (Fig. 4A, left panel). Surprisingly, there seemed to be more cyclin-D1-positive cells in the T-treated HBT than in the control HBT, and few cyclin-D1expressing cells were found in the DHT-treated explants. The qRT-PCR results showed that DHT decreased cyclin-D1 mRNA (Fig. 4B), whereas the level of cyclin-D1 mRNA was increased in T-treated HBT.
ever, the level of cyclin-D1 mRNA in the T-treated HBT was much lower than that in the E2-treated HBT. In the combination cultures, both androgens clearly decreased E2-induced cyclin-D1 mRNA (Fig. 4C).

Androgens increase apoptosis in cultured HBT

To investigate how androgen signaling affects cell survival in HBT, cleaved caspase-3 (CPP32), a marker for apoptosis, was assessed (Fig. 5). The proportion of CPP32-positive cells was increased in the HBT cultured with T (P < 0.001) and DHT (P < 0.001) for 7 and 14 d compared with the controls (Fig. 5). E2/T and E2/DHT reduced the percentage of apoptotic cells significantly when compared with the control and E2-alone groups.
Expression of AR, ERα, and ERβ in androgen- and E2-treated HBT

Immunohistostainings showed that in noncultured HBT, 34% of the cells were positive for the AR, 26% for ERα, and 80% for ERβ. The AR were mainly located in the nuclei of acinar and ductal epithelial cells and, to a minor extent, in stromal cells. ERα was observed only in epithelial cells, and ERβ was observed in the nuclei of epithelial cells and in scattered stromal cells. The percentage of AR cells was significantly increased in the T and DHT groups compared with the controls, whereas E2 treatment reduced the proportion of AR cells (Fig. 6A). Addition of E2 to T and DHT raised AR-positive cell numbers compared with E2-alone treatment. The count of ERα-positive cells was lowered by T and DHT in comparison with the control (P < 0.001), whereas E2 increased them significantly (Fig. 6B). The E2 effect was opposed by T and DHT in the E2/T and E2/DHT cultures (Fig. 6B). The proportion of ERβ-positive cells was not affected by T and DHT, but E2 significantly decreased their expression (Fig. 6C). The combination of T and DHT with E2 opposed the E2 effects, but the number of ERβ-positive cells remained lower than in the controls.

Discussion

In this study, we investigated how androgens influence HBT and how they modulate E2 stimulation of the breast. We used organ cultures of HBT obtained from reduction mammoplasty surgeries of postmenopausal women in which HBT may differ from breast tissues in other populations. As we reported previously, the HBT explants retain their structural integrity and appropriate hormonal responses in this model (13).

The maintenance of hormone responsiveness of cultured HBT was demonstrated by studying the expression of known androgen and E2 target genes (14, 18). PSA is widely used as an androgen-regulated marker in prostate tissue, but it is also expressed in the breast (19) and other tissues (20). Our cultures showed that T and DHT strongly induced PSA expression in HBT. Bclt inhibition of androgen-induced PSA expression demonstrated that expression was mediated by the AR. The percentage of PSA-positive cells in the E2, Fulv, and E2/Fulv cultures decreased, the decrease of which was probably caused by indirect mechanisms such as repression of AR expression (21) and because the PSA gene is not known to contain estrogen response elements (14). Extranuclear ER actions (22) provide another possible mechanism for E2 suppression of PSA. The expression of Apo-D, another androgen target gene (14), was also induced in an AR-dependent manner. These results suggest that T and DHT can directly influence target gene expression in HBT.

The strong up-regulation of TFF1 and AREG expression in the presence of E2 showed the appropriate response to E2 and Fulv inhibition of stimulation indicated an ER-mediated effect. Fulv alone decreased TFF1 mRNA, suggesting that TFF1 expression in basal conditions may have been slightly increased by ER activation by endogenous ligands (a possibility discussed later) or by ligand-independent mechanisms. Both androgens inhibited TFF1 expression. Interestingly, Bclt alone and T/Bclt increased TFF1 mRNA compared with the control. This finding suggests that the AR is involved in the repression of TFF1 expression or in decreasing the proportion of cells expressing the protein. It is also possible that T aromatization to E2 may contribute to increased TFF1 mRNA level in T/Bclt group.

The E2 stimulation of epithelial cell proliferation is generally considered to increase the risk for the development of breast cancer (23). However, conflicting studies have been reported on the effects of exogenous androgens on the proliferation of mammary epithelial cells and the as-
associated risk of breast cancer (5, 6, 24–28). Our study demonstrates that androgens decreased proliferation in a time-dependent manner and effectively opposed the stimulatory effects of E2. Interestingly, Bclt increased the proportion of proliferating cells, which suggests that AR exerts an inhibitory effect on the basal levels of proliferation. Our results agree with a previous study on human breast tissue by Hofling et al. (29), who found decreased Ki67 cells in women treated with androgen-containing hormone therapy. Corresponding results have also been obtained in primate model studies in which androgen treatment decreased mammary proliferation (11).

The mechanisms of androgen inhibition of HBT proliferation were further evaluated by studying the expression of cyclin-D1, which has a crucial role in triggering cell cycle progression from the G1 to the S phase. In our experiments, DHT decreased cyclin-D1 mRNA and immunostaining. This result agrees with earlier observations with breast cancer cells (30). Comstock et al. (31) recently reported that ligand-bound AR interacts with specific androgen response element sequences on the cyclin-D1 promoter, which leads to the inhibition of cyclin-D1 transcription (30). In contrast, T increased cyclin-D1 mRNA levels. This effect may partly have been caused by androgen conversion to E2 (10), as discussed later. Unexpectedly, Bclt alone and together with T or DHT decreased cyclin-D1 mRNA, which suggests promoter-specific Bclt effects and generally reflects the complex actions of ligand-bound AR on the cyclin-D1 promoter (30). The overall effects of T were antiproliferative, opposed by Bclt, which suggests that the regulation of cyclin-D1 was not critical among the mechanisms leading to the T-induced inhibition of proliferation. In combination cultures with E2, both T and DHT clearly opposed the strong up-regulation of cyclin-D1 expression by E2.

In addition to inhibiting proliferation, androgens contributed to epithelial regression by increasing apoptosis, an effect counteracted by E2 in the combination culture. DHT has been reported to induce cell death (32) and inhibit the expression of antiapoptotic bcl-2 in breast cancer cells (8, 33), which could be a mechanism for increased cell death in androgen-treated HBT. However, in vivo treatment with DHT arrested growth but did not induce cell death in developing mouse mammary glands (34).
Androgens, T, and nonaromatizable DHT exerted similar effects on the proliferation, apoptosis, and expression of the marker proteins. As expected, DHT was generally more potent than T. In premenopausal women, circulating androgens include both T and DHT. In postmenopausal women, androgens are primarily formed from adrenal precursors in peripheral tissues, such as the breast (35). The enzymatic activities for steroid sulfatases and 5α-reductase have been demonstrated in normal HBT (1), which allow the synthesis of T and the conversion of T to DHT in breast tissue. In our HBT cultures, the T effects could be partly mediated by the more bioactive DHT. As
previously reported (36), HBT was found to express low levels of aromatase catalyzing the conversion of T to E2 (Supplemental Fig. 1). As discussed earlier, the conversion of T to E2 may have contributed to T-increased cyclin-D1 mRNA expression, which was inhibited by DHT. Owing to lack of feasible methods, concentrations of steroids and their metabolites in small explants could not be measured, but according to previous reports, their levels in normal postmenopausal HBT are very low (37). The overall similarity of other T and DHT responses and the experiments with Bclt suggest that the majority of the effects of T and DHT on proliferation and apoptosis in breast epithelial cells are mediated by the AR. Thus, the local production of androgens and the metabolism of T and E2 may not have played a major role in culture conditions.

Immunohistochemistry showed that E2 and androgen treatment markedly altered the percentage of ERα- and AR-positive cells in mammary epithelium. The androgen treatment increased the proportion of AR-positive cells, whereas ERα-expressing cells were strongly decreased. The ERβ-expressing cells remained unaltered. Treatment with E2 increased the number of ERα-positive cells, whereas those of ERβ- and AR-positive cells decreased. In the combination cultures, E2 and the androgens opposed each other. The altered proportions of steroid receptor-positive cells in the E2 and androgen treatments could be explained by changes in receptor gene expression and/or the enrichment or reduction of receptor-positive cells, due to changes in proliferation, and/or apoptosis of selected cell populations. DHT has been found to increase AR expression (30) and decrease ERα expression in primate mammary glands in vivo (11) and in human breast cancer cells (8, 30). Our present experiments do not reveal the extent to which these mechanisms are involved in the changes in AR- and ER-positive cell percentages. The altered proportions of AR- and ER-expressing cells have an important role in E2 stimulation and androgen repression of proliferation in mammary epithelial cells and in mammary gland response to hormonal therapies. Accordingly, one mechanism that could explain the androgen inhibition of proliferation is a strong decrease in ERα-positive cells. Whether AR-positive cells are able to divide in the mammary epithelium remains unknown, but recent reports demonstrated that androgens inhibit cyclin-D1 expression in MCF-7 cells in an AR-
dependent manner, which has been considered an explanation for androgen inhibition of proliferation in these cells (30).

An increase in ERα-positive cells was associated with stimulated proliferation and decreased apoptosis in E2-treated explants. Conversely, decreased ERα positivity was linked to decreased proliferation and increased apoptosis in androgen-treated cultures. Previous studies (38) have shown that ERα-positive mammary epithelial cells do not proliferate, whereas E2-induced growth is mediated by paracrine stimulation of division in adjacent ERα-negative cells (38). A critical mediator of ER action is AREG (16), which was induced by E2 in our HBT cultures. Because ERα-positive cells increased in the presence of E2, it is likely that ER-negative cells rapidly and increasingly differentiate to ERα-positive cells after cell division.

Interestingly, the response of ERβ-positive cell numbers to the T, DHT, and E2 treatments seemed to resemble those of AR-positive cells, whereas the responses of ERα- and ERβ-expressing cells were mostly the opposite. The function of ERβ in breast tissue is largely unknown, but it is thought to have protective functions, similar to AR (39). Correspondingly, the expression of AR and ERβ in tumor specimens has been associated with favorable outcomes in breast cancer patients (39).

Our results demonstrate that in addition to their direct inhibitory effects, T and DHT oppose the effects of E2 on proliferation, apoptosis, and target gene expression in HBT, as previously reported in various experimental in vivo and in vitro models (6, 11). The mechanisms by which androgens inhibit E2 in breast tissue may include T- and DHT-induced decreases in ERα-positive cells and/or in ERα protein expression (11, 30) or increased apoptosis of ERα-positive cells. Peters et al. (40) recently suggested that AR strongly inhibits ERα activity by binding to estrogen response elements in a subset of ERα target genes, thereby preventing ER activation of these genes. Another mechanism that was recently suggested for maintaining the balance between ER/AR actions is the suppression of AR target gene expression by cyclin-D1 (31). In addition to controlling the cell cycle, cyclin-D1 has other functions including the modulation of nuclear receptor actions (31). Comstock et al. (31) recently reported that cyclin-D1 can block the binding of AR to a subset of target genes, thereby inhibiting androgen action. If corresponding mechanisms work in the mammary epithelium, then E2-induced cyclin-D1

**FIG. 6.** The expression of AR, ERα, and ERβ in noncultured and cultured HBT. Photomicrographs show the immunoreactivity for AR (A), ERα (B), and ERβ (C) in noncultured HBT. Scale bar, 100 μm. The scores of AR-, ERα-, and ERβ-positive cells were calculated as percentages of positive cells. The columns show the mean ± SD values (n = 12) for the AR-, ERα-, and ERβ-immunopositive cells in the different treatment groups. The asterisks (*, P < 0.05; **, P < 0.01; ***, P < 0.001) indicate the statistical significance of differences between the treatment and control group.
could effectively inhibit androgen regulation of selected target genes and their functions.

Normal breast tissue is exposed to considerable levels of androgens (35). In premenopausal women, they are circulating androgens, such as T and DHT. In postmenopausal women, these androgens are primarily formed from adrenal precursors in peripheral tissues (10, 35). Thus, androgens belong to the physiological environment of normal breast tissue, and they may effectively balance the stimulatory effects of estrogens (1). E2 and E2/progesterone treatments in postmenopausal women lead to decreased serum androgen levels by feedback mechanisms and other systemic effects, such as changes in SHBG production. This disrupts the balance between hormones and may contribute to a risk of excessive E2 stimulation of the breast (1). Therefore, adding androgen to postmenopausal hormone therapy has been suggested (6). In clinical use, androgen treatment for hypoactive sexual desire is associated with several beneficial effects in postmenopausal women, such as improved sexual function, general well-being and positive effects on bone mineral density (10, 41). However, androgen use causes side effects, including virilizing effects (acne and hirsutism), adverse lipid changes, reduced insulin sensitivity, and an altered pattern of fat deposition (10, 41). High levels of serum androgens have also been suggested to provide increased precursor supplies for E2 synthesis in aromatase reactions, leading to the stimulation of proliferation in breast and even increased risk of breast cancer (9). Considering the strong inhibition of E2 effects by androgens, this mechanism may not markedly affect the safety of androgen therapy at the tissue level. The development of novel selective androgen receptor modulators, which retain antiproliferative effects on mammary epithelium but lack adverse effects, could provide an important component to breast protective hormone therapy in the future.

Acknowledgments

We thank Mr. Ioan Iagar for technical support, Ms. Soili Jussila for help with immunostaining assays, Mrs. Liudmila Shumskaya for preparation of the paraffin sections of tissues, and the personnel of the Department of Surgery (Turku University Hospital, Turku, Finland) for generously providing samples.

Address all correspondence and requests for reprints to: Pirkko Härkönen, M.D., Ph.D. (member of The Endocrine Society, membership no. 140128), Department of Cell Biology and Anatomy, Institute of Biomedicine, University of Turku, Kimanymyllynkatu 10, 20520 Turku, Finland. E-mail: harkonen@utu.fi.

This work was supported by Turku University Hospital and the Sigrid Jusélius and Turku University Foundation. The fund-

References


