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Published in:
PLoS ONE

DOI:
10.1371/journal.pone.0071846

2013

Link to publication

Citation for published version (APA):

Total number of authors:
12

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Plasma Lipid Composition and Risk of Developing Cardiovascular Disease

Celine Fernandez1*, Marianne Sandin2, Julio L. Sampaio3, Peter Almgren1, Krzysztof Narkiewicz4, Michal Hoffmann4, Thomas Hedner5, Björn Wahlstrand5, Kai Simons3, Andrej Shevchenko3, Peter James2, Olle Melander1*

1 Department of Clinical Sciences, Lund University, Malmö, Sweden, 2 Department of Immunotechnology, Lund University, Lund, Sweden, 3 Max Planck Institute of Molecular Cell Biology and Genetics, Dresden, Germany, 4 Department of Hypertension and Diabetology, Medical University of Gdansk, Gdansk, Poland, 5 Department of Medicine, Sahlgrenska Academy, Göteborg University, Göteborg, Sweden

Abstract

Aims: We tested whether characteristic changes of the plasma lipidome in individuals with comparable total lipids level associate with future cardiovascular disease (CVD) outcome and whether 23 validated gene variants associated with coronary artery disease (CAD) affect CVD associated lipid species.

Methods and Results: Screening of the fasted plasma lipidome was performed by top-down shotgun analysis and lipidome compositions compared between incident CVD cases (n = 211) and controls (n = 216) from the prospective population-based MDC study using logistic regression adjusting for Framingham risk factors. Associations with incident CVD were seen for eight lipid species (0.21≤q≤0.23). Each standard deviation unit higher baseline levels of two lysophosphatidylcholine species (LPC), LPC16:0 and LPC20:4, was associated with a decreased risk for CVD (P = 0.024–0.028). Sphingomyelin (SM) 38:2 was associated with increased odds of CVD (P = 0.057). Five triglyceride (TAG) species were associated with protection (P = 0.031–0.049). LPC16:0 was negatively correlated with the carotid intima-media thickness (P = 0.010) and with HbA1c (P = 0.012) whereas SM38:2 was positively correlated with LDL-cholesterol (P = 0.06–0.07) and the q-values were good (q≤0.03). The risk allele of 8 CAD-associated gene variants showed significant association with the plasma level of several lipid species. However, the q-values were high for many of the associations (0.015≤q≤0.75). Risk allele carriers of 3 CAD-loci had reduced level of LPC16:0 and/or LPC 20:4 (P≤0.056).

Conclusion: Our study suggests that CVD development is preceded by reduced levels of LPC16:0, LPC20:4 and some specific TAG species and by increased levels of SM38:2. It also indicates that certain lipid species are intermediate phenotypes between genetic susceptibility and overt CVD. But it is a preliminary study that awaits replication in a larger population because statistical significance was lost for the associations between lipid species and future cardiovascular events when correcting for multiple testing.

Introduction

Cardiovascular mortality and morbidity is a major public health problem in Western societies. Traditional cardiovascular risk factors do not fully explain future cardiovascular events [1,2] and adding modern biomarkers to the standard risk factors has, thus far, only proven to minimally improve individual risk prediction [3,4], thus underlining the need to identify new biomarkers.

Lipids are thought to play a central role in cardiovascular disease (CVD) development and total plasma triglycerides and cholesterol as well as LDL- and HDL-cholesterol are traditionally monitored as predictors of cardiovascular events. However, those are crude measurements of the sum of a complex composition of lipids and do not at all reflect other potentially atherogenic lipid species. We here hypothesized that specific plasma lipid species, rather than the rough phenotype of total triglycerides and cholesterol may be altered in subjects who develop CVD later in life, implying that they may be involved in the CVD pathogenesis.

Lipidomics, a subset within the field of metabolomics, strives to quantitatively describe the complete set of all lipids in a given cell type, tissue or biologic fluid of interest at a given time [5]. There is no single instrument or approach that can currently do so, but
Materials and Methods

Materials, Chemicals and Lipid Standards

Material resistant to organic solvent was used (e.g. polypropylene, silicone, Teflon). Synthetic lipid standards were purchased from Avanti Polar Lipids, Inc. (Alabaster, AL) or Larodan Fine Chemicals (Malmö, Sweden). Methyl-tert-butylether (MTBE) and Acetone were purchased from Sigma-Aldrich (Munich, Germany). Methanol, chloroform and ammonium acetate (Liquid Chromatography grade) were purchased from Fluka (Buchs SG, Switzerland) and 2-propanol (ACS grade) from Sigma-Aldrich (Munich, Germany).

Table 1. Baseline characteristics of the study samples.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Control (n = 216)</th>
<th>CVD case (n = 211)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>60.7 ± 5.1</td>
<td>60.2 ± 5.3</td>
<td>0.331</td>
</tr>
<tr>
<td>Women (%)</td>
<td>47.7</td>
<td>47.4</td>
<td>0.952</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>26.3 ± 4.3</td>
<td>26.5 ± 4.4</td>
<td>0.558</td>
</tr>
<tr>
<td>Systolic blood pressure (mm Hg)</td>
<td>149.3 ± 19.8</td>
<td>149.7 ± 18.4</td>
<td>0.815</td>
</tr>
<tr>
<td>Diastolic blood pressure (mm Hg)</td>
<td>90.1 ± 9.6</td>
<td>90.1 ± 9.3</td>
<td>0.981</td>
</tr>
<tr>
<td>Glucose (mmol/l)</td>
<td>5.2 ± 1.2</td>
<td>5.7 ± 2.2</td>
<td>0.008</td>
</tr>
<tr>
<td>Cholesterol (mmol/l)</td>
<td>6.3 ± 1.1</td>
<td>6.3 ± 1.0</td>
<td>0.611</td>
</tr>
<tr>
<td>Triglycerides (mmol/l)</td>
<td>1.5 ± 0.7</td>
<td>1.4 ± 0.6</td>
<td>0.136</td>
</tr>
<tr>
<td>High density lipoprotein (mmol/l)</td>
<td>1.3 ± 0.3</td>
<td>1.3 ± 0.3</td>
<td>0.541</td>
</tr>
<tr>
<td>Low density lipoprotein (mmol/l)</td>
<td>4.3 ± 1.0</td>
<td>4.4 ± 1.0</td>
<td>0.426</td>
</tr>
<tr>
<td>Diabetes (%)</td>
<td>8.3</td>
<td>15.2</td>
<td>0.028</td>
</tr>
<tr>
<td>Current smoker (%)</td>
<td>33.3</td>
<td>33.6</td>
<td>0.945</td>
</tr>
<tr>
<td>Anti-hypertensive treatment (%)</td>
<td>25.9</td>
<td>23.7</td>
<td>0.594</td>
</tr>
<tr>
<td>Lipid lowering drugs (%)</td>
<td>0.9</td>
<td>3.8</td>
<td>0.050</td>
</tr>
</tbody>
</table>

Values are mean ± s.d. or percentage. P values were calculated using a t test for continuous variables and Pearson Chi-Square for binary variables. doi:10.1371/journal.pone.0071846.t001
methanol 5/1.5 (v/v) providing a total of 2.7 nmol cholesteryl heptadecanoate (CE17:0), 0.7 nmol heptadecanoyl sphingomyelin (SM17:0), 3.5 nmol 1,2-di-O-hexadecyl-sn-glycero-3-phosphocholine (PC-O12:0/−O12:0), 0.9 nmol 1,2-di-O-phytanoyl-sn-glycero-3-phosphoethanolamine (PE-O16:0/−O16:0), 3.1 nmol 1-lauroyl-2-hydroxy-sn-glycero-3-phosphocholine (LPC12:0), 0.4 nmol N-heptadecanoyl-D-mythro-sphingosine (Cer17:0), 3.1 nmol tri- laurin (TAG12:0) and 0.5 nmol dilaurin (DAG12:0) ). Then 350 μL of MTBE/methanol 5:1.5 were added and the samples were shaken at 4°C for 1 h. Afterwards, 150 μL of water were added, followed by shaking at 4°C for 10 min and centrifugation for 5 min at 4,000 rpm on Rotanta 460R centrifuge (Hettich, Tuttlingen, Germany). The upper organic phase was transferred into a 96-well plate with glass inserts and a silicone/Teflon coated sealing mat (Chromacol) and stored at −20°C until performing the MS analysis for all the samples successively.

**Shotgun Screening of Plasma Lipidome**

Prior to the MS analysis, the lipid extracts were diluted 10 times with a mixture of chloroform/methanol/2-propanol 1/2/4 (v/v/v) containing 7.5 mM ammonium acetate and placed in a 96-well plate (Eppendorf) that was then sealed with aluminium foil (Corning). Shotgun analysis was performed on a LTQ Orbitrap (Thermo Fisher Scientific, Waltham, MA) coupled to a TriVersa NanoMate robotic nanoflow ion source (Advion BioSciences, Ithaca, NY) [8,12]. Samples were analyzed in duplicate. Lipids were identified and quantified using the LipidXplorer software [24] and lipid species of the following lipid classes were recognized: triacylglyceride (TAG), diacylglyceride (DAG), cholesteryl ester (Chol-FA), sphingomyelin (SM), phosphatidylcholine (PC), PC-ether (PC-O), lyso-PC (LPC), phosphatidylethanolamine (PE) and PE-ether (PE-O). Identification of the different lipid species was based on MS survey scans acquired in positive ion mode in the Orbitrap analyzer at a target mass resolution of 100,000 using a mass accuracy of better than 5 ppm and a signal to noise ratio of 2. Lipid species were quantified by normalizing the intensities of their peaks to the intensity of the peaks of internal standards spiked into the sample prior to lipid extraction. The internal standards were also used to monitor the quality of the MS analysis and representative mass spectra are presented (Supplementary Figure S1A and S1B). An internal standard mix was both extracted and run independently 18 times across the entire analysis to get an estimate of the coefficient of variation of the combined lipid extraction and MS analysis from the internal standards (Supplementary Table S1). The maximum value of duplicate samples was kept. Lipid species with >30% missing observations were excluded.

**Statistical Analyses**

SPSS (version 18.0) was used for all statistical analyses. Data were assessed for normality with histograms. Due to non-normality all the lipid species were log transformed prior analysis. All tests were two-sided and data were considered significant if P<0.05.

To determine the association of baseline individual lipid species with future CVD, we performed binary logistic regression adjusting for age, sex, diabetes, smoking status, LDL-cholesterol, HDL-cholesterol, systolic blood pressure (SBP), body mass index (BMI) and use of anti-hypertensive treatment.

Q-values were calculated using the QVALUE software [25]. Hierarchical clustering was performed with Euclidean distance and average linkage in MATLAB R2011a (version 7.12.0.635).
Results

Lipid Metabolites Profiling in the Cardiovascular Cohort of the Malmö Diet and Cancer Study

As a result of the initial matching procedure (age, gender and Framingham risk score) the baseline characteristics of the 211 incident cases of CVD and 216 control subjects were similar for most risk factors except fasting plasma glucose level and diabetes. The frequency of use of lipid lowering drugs was low (Table 1).

Lipid profiling was performed on samples obtained from the baseline examination that took place between 1991 and 1994. A total of 85 lipid species belonging to 9 major lipid classes were identified and quantified by the approach used (Supplementary Table S2). The total quantities of triglycerides and cholesterol determined by mass spectrometry were correlated with the values obtained by traditional clinical chemistry analysis (Figure 1 and Supplementary Figure S2). As known from previous study, the correlation was substantially stronger for triglycerides than for cholesterol [8].

Selected Lipid Species Associate with Future Adverse Cardiovascular Disease Outcome

Binary logistic regression was performed to assess the association between baseline lipid species level and future CVD adjusting for Framingham risk factors. Associations with incident CVD were seen for lipid species belonging to the lysophosphatidylcholine (LPC), sphingomyelin (SM) and triacylglyceride (TAG) lipid classes, but the q-values for the associations were rather high (0.21 ≤ q ≤ 0.23) (Tables 2, 3 and Supplementary material online, Table S3A). Similar results were obtained when only adjusting for diabetes (Supplementary Table S3B).

In the LPC class, each standard deviation (SD) unit higher baseline levels of LPC16:0 or LPC20:4 was associated with a decreased risk of developing CVD over the 12-year follow-up period (OR = 0.79; P = 0.028 and OR = 0.77; P = 0.024, respectively) (Table 2). Individuals whose plasma level of LPC16:0 or LPC20:4 was in the top quartile had decreased odds of future CVD compared with individuals in the lowest quartile (OR = 0.57; P = 0.032 and OR = 0.62; P = 0.048, respectively) (Table 2).

SM38:2, with a borderline P-value, was the only lipid species of its class to be associated with increased odds of future CVD.

### Table 2. Relation of baseline phospholipids level to future adverse cardiovascular outcome adjusting for Framingham risk factors.

<table>
<thead>
<tr>
<th>Model</th>
<th>LPC16:0 (n = 424)</th>
<th>LPC20:4 (n = 353)</th>
<th>SM38:2 (n = 318)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Models adjusting for sex, age, BMI, type 2 diabetes, anti-hypertension treatment, smoking, LDL, HDL and SBP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lipid species as continuous variable</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Per s.d.</td>
<td>0.79 (0.65–0.97)</td>
<td>0.77 (0.61–0.96)</td>
<td>1.28 (0.99–1.64)</td>
</tr>
<tr>
<td>p-value</td>
<td>0.028</td>
<td>0.024</td>
<td>0.057</td>
</tr>
<tr>
<td>q-value</td>
<td>0.210</td>
<td>0.210</td>
<td>0.228</td>
</tr>
<tr>
<td>Lipid species as categorical variable</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>First quartile</td>
<td>1.0 (referent)</td>
<td>1.0 (referent)</td>
<td>1.0 (referent)</td>
</tr>
<tr>
<td>Second quartile</td>
<td>1.21 (0.69–2.11)</td>
<td>1.13 (0.61–2.07)</td>
<td>0.94 (0.49–1.81)</td>
</tr>
<tr>
<td>Third quartile</td>
<td>0.94 (0.54–1.65)</td>
<td>0.62 (0.34–1.16)</td>
<td>1.32 (0.68–2.56)</td>
</tr>
<tr>
<td>Fourth quartile</td>
<td>0.57 (0.32–1.00)</td>
<td>0.62 (0.33–1.17)</td>
<td>1.85 (0.92–3.71)</td>
</tr>
<tr>
<td>p for trend</td>
<td>0.032</td>
<td>0.048</td>
<td>0.054</td>
</tr>
</tbody>
</table>

Values are odds ratios (95% confidence intervals) for cardiovascular disease from multivariate adjusted binary logistic regressions performed with the Z score of a given lipid species obtained after log transformation. BMI, body mass index; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; LPC, lysophosphatidylcholine; SBP, systolic blood pressure; SM, sphingomyelin.

doi:10.1371/journal.pone.0071846.t002

### Table 3. Relation of baseline triglycerides level to future adverse cardiovascular outcome adjusting for Framingham risk factors.

<table>
<thead>
<tr>
<th>Model</th>
<th>TAG48:1 (n = 424)</th>
<th>TAG48:2 (n = 424)</th>
<th>TAG48:3 (n = 402)</th>
<th>TAG50:3 (n = 424)</th>
<th>TAG50:4 (n = 423)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Models adjusting for sex, age, BMI, type 2 diabetes, anti-hypertension treatment, smoking, LDL, HDL and SBP</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lipid species as continuous variable</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Per s.d.</td>
<td>0.78 (0.63–0.98)</td>
<td>0.79 (0.64–0.98)</td>
<td>0.81 (0.65–1.00)</td>
<td>0.79 (0.63–0.98)</td>
<td>0.79 (0.64–0.98)</td>
</tr>
<tr>
<td>p-value</td>
<td>0.031</td>
<td>0.034</td>
<td>0.049</td>
<td>0.036</td>
<td>0.033</td>
</tr>
<tr>
<td>q-value</td>
<td>0.210</td>
<td>0.210</td>
<td>0.228</td>
<td>0.210</td>
<td>0.210</td>
</tr>
</tbody>
</table>

Values are odds ratios (95% confidence intervals) for cardiovascular disease from multivariate adjusted binary logistic regressions performed with the Z score of a given triacylglyceride species obtained after log transformation. BMI, body mass index; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; SBP, systolic blood pressure; TAG, triacylglyceride.

doi:10.1371/journal.pone.0071846.t003
Table 4. LPC16:0 and LPC20:4 negatively correlate with CVD risk factors whereas SM38:2 positively correlates with CVD risk factors.

<table>
<thead>
<tr>
<th>Lipid specie</th>
<th>LPC16:0</th>
<th>LPC20:4</th>
<th>SM38:2</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Correlation</td>
<td>p-value</td>
<td>q-value</td>
</tr>
<tr>
<td>Imtcca0</td>
<td>-0.13</td>
<td>0.010</td>
<td>0.007</td>
</tr>
<tr>
<td>HbA1c</td>
<td>-0.12</td>
<td>0.012</td>
<td>0.009</td>
</tr>
<tr>
<td>BMI</td>
<td>-0.14</td>
<td>0.004</td>
<td>0.003</td>
</tr>
<tr>
<td>SBP</td>
<td>-0.06</td>
<td>0.244</td>
<td>0.114</td>
</tr>
<tr>
<td>LDL</td>
<td>0.07</td>
<td>0.154</td>
<td>0.081</td>
</tr>
<tr>
<td>HDL</td>
<td>0.12</td>
<td>0.014</td>
<td>0.011</td>
</tr>
</tbody>
</table>

Partial correlations were performed between LPC16:0, LPC20:4, or SM38:2 after log transformation and current known laboratory predictors for cardiovascular disease, adjusting for age and sex. BMI, body mass index; HbA1c, haemoglobin A1c; HDL, high-density lipoprotein cholesterol; Imtcca0, intima-media thickness of the common carotid artery at baseline; LDL, low-density lipoprotein cholesterol; LPC, lysophosphatidylcholine; SBP, systolic blood pressure; SM, sphingomyelin.

Discussion

Top-down Lipidomics, a Tool for Clinical Screens

The importance of two main lipids, i.e. triglycerides and cholesterol, as a tool for CVD prediction has long been known. But, modern lipidomics analysis shows that the human plasma lipidome comprises of at least several hundreds of individual lipid species and gives a glimpse of the complexity of the lipidome that has been overlooked until recently mainly because of technical limitations. We here performed a plasma lipidome screen in a prospective population-based cohort using top-down shotgun lipidomics. We aim to look for differences in the plasma composition in individuals with similar plasma total lipids level. We analyzed 427 samples with 2 technical replicates and identified and quantified 85 lipid species belonging to 9 different lipid classes and to our knowledge this study constitutes the first extensive lipid profiling of plasma for incident CVD in the primary preventive setting. Top-down shotgun lipidomics was the method of choice for this study because it is a quantitative and highly sensitive technique that allows high-throughput and relatively extensive lipid coverage.
Figure 2. Association between the lipid profile and the risk allele of 8 CAD-associated gene variants. Heat map of regression coefficients obtained from linear regressions performed between the CAD-associated locus (with the CAD-associated allele coded) and the lipid species after log transformation adjusting for age and sex. *P<0.05, **P<0.01, ***P<0.001.
doi:10.1371/journal.pone.0071846.g002
Refining the Dyslipidemia Phenotype

Although total increased plasma TAG concentration is considered a risk factor for CVD, we have here identified some individual TAG species that were associated with decreased odds of future CVD. LPC as a whole lipid class has previously been linked with inflammation as well as with both pro- and anti-atherogenic effects [26,27], whereas we have here shown that two specific LPC species i.e., LPC16:0 and LPC20:4, were protective for CVD. These findings demonstrate that systematic analysis of plasma lipid species rather than lipid classes as a whole may reveal opposite relationships with CVD risk and thus could help to better understand the mechanisms leading to CVD and to improve CVD risk prediction.

Recently, a plasma lipidomics analysis was conducted using a different MS platform than the one we used, in a cross sectional setting, showing that certain patterns of lipids could discriminate between patients with stable angina and those with unstable CAD as well as healthy controls [9]. The results obtained in patients with unstable CAD are supported and extended by our prospective study of subjects without prior CVD, i.e., decreased level of most measured LPC species both in CAD versus control as well as in unstable versus stable CAD, increased levels of several SM species in unstable versus stable CAD and decreased level of specific TAG species in unstable versus stable CAD, were reported. This suggests that such alterations of lipid patterns may not only be a marker of coronary atherosclerosis and plaque instability but also that it may play a role in the pathogenesis of CVD, given its presence more than 10 years before clinical disease onset.

Integrating Genomic and Lipidomics Information

Out of the 8 CAD susceptibility gene variants displaying significant association with circulating lipid species concentrations, 3 have not yet been previously reported to be involved in lipid metabolism (WDR12, ZC3HC1 and PHACTR1) and 3 are only known to affect lipoproteins levels (LPA, SORT1 and the ZNF259/APOA5-A4-C3-A1 gene region) [13,28]. However, any potential link between the genetic alteration of these lipids and CAD needs to be substantiated by mechanistic studies. Two of the 8 CAD loci are directly coding for enzymes involved in lipids biosynthesis (PPAP2B and the PEMT/RASD1/SMCR3 locus) [29,30]. The PPAP2B gene encodes a phosphatidic phosphatase that converts phosphatidic acid into diacylglycerol, the precursor for de novo synthesis of TAG, PC and PE. Moreover, PEMT encodes an enzyme which sequentially converts PE into PC. Both carriers of the PPAP2B and of the PEMT/RASD1/SMCR3 risk allele display reduced level of multiple glycerophospholipids including the CVD-protective lipid species LPC16:0 and/or LPC20:4. Overall, our findings highlight that integrating lipidomics with genomics is a promising approach to increase the understanding of mechanisms underlying the gene-CVD associations as well as CVD pathogenesis.

Study Limitations

This is an initial discovery study that needs to be replicated especially since the false discovery rate was high when looking for associations between the lipid species and future cardiovascular events or between the lipid species and most of the CAD-associated gene variants. Also, we do acknowledge that this is a case control study and not a general population study, thus the findings cannot be generalised to the whole population. Furthermore, our study could be complemented by acquiring spectra in negative ion mode to extend the lipid class coverage and by performing tandem MS for some targeted lipid species in order to get their full structural information. Another draw-back of the study is the lack of a pooled quality control plasma sample run across the study. Finally, we do not know to what extent the −80 degree Celsius storage over approximately 20 years may have affected the original lipid profile.

Conclusions

This study constitutes a proof-of-concept screen that shotgun lipidomics can be used as a tool in the search for novel CVD biomarkers. Moreover, we here highlight the importance of refining the dyslipidemia phenotype and thus looking at the level of individual lipid species rather than the total sum of the different lipid classes in their relationship with CVD risk. We identified some specific lipid species as potential biomarkers of adverse cardiovascular outcome. However, statistical significance was lost for the association between the lipid species and future cardiovascular events when correcting for multiple testing. Finally, our results support the informative value in bringing together genomic and lipidomics data, suggesting that certain individual lipid species are intermediate phenotypes between genetic susceptibility and overt CVD. Overall, this is an explorative study that will need to be replicated in a larger population.

Supporting Information

Figure S1 Representative mass spectra of total lipid extracts from plasma. The most abundant peaks are annotated with m/z; the shaded areas indicate the m/z ranges where the corresponding lipid classes were detected.

Figure S2 Absolute quantification of TAGs by top-down lipidomics correlates with the total triglyceride levels measured at baseline examination. Linear regression was performed between the total absolute TAG levels determined by MS versus the total triglyceride levels measured by traditional clinical chemistry analysis. The total TAG level measured by MS is obtained by summing the abundances of all the individual TAG species.

Table S1 Coefficient of variation (CV) of the combined lipid extraction and MS analysis for the 8 internal standards.

Table S2 Absolute levels of the lipid species.

Table S3 A. Relation of baseline lipid species level to future adverse cardiovascular outcome adjusting for Framingham risk factors. B. Relation of baseline lipid species level to future adverse cardiovascular outcome adjusting for type 2 diabetes only.

Table S4 Relation of baseline triglycerides species level to future adverse cardiovascular outcome adjusting for Framingham risk factors.
Table S5  Estimated q-values of the tests performed to study the association between CVD risk factors and the lipid species.

Table S6  Relation between 23 validated coronary artery disease associated gene variants and baseline plasma lipid metabolites level.

Table S7  The risk allele of 8 of the validated coronary artery disease associated gene variants shows significant association with the baseline plasma level of several lipid species.

References


